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(54) Title: GOODPASTURE ANTIGEN BINDING PROT (57) Abstract The present invention provides isolated nucleic acid protein (GPBP), substantially purified GPBP, antibodies again	l segue	ces and expression vectors encoding the Goodpasture antigen binding BP, and methods for detecting GPBP.			

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GOODPASTURE ANTIGEN BINDING PROTEIN

Cross Reference

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This application claims priority to U.S. Provisional Patent Application Serial No. 60/121,483, filed February 24, 1999.

Statement of Government Rights

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15 Field of the Invention

The invention relates to the fields of protein kinases, automimmune disease, apoptosis, and cancer.

Background of the Invention

Goodpasture (GP) disease is an autoimmune disorder described only in humans. In GP patients, autoantibodies against the non-collagenous C-terminal domain (NC1) of the type IV collagen $\alpha 3$ chain ("Goodpasture antigen") cause a rapidly progressive glomerulonephritis and often lung hemorrhage, the two cardinal clinical manifestations of the GP syndrome (see 1 for review. The reference numbers in this section correspond to reference list of Example 1).

The idea that common pathogenic events exist at least for some autoimmune disorders is suggested by the significant number of patients displaying more than one autoimmune disease, and also by the strong and common linkage that some of these diseases show to specific MHC haplotypes (31, 32). The experimental observation that the autoantigen is the leading moiety in autoimmunity and that a limited number of self-components are autoantigenic (31), suggest that these self-components share biological features with important consequences in self/non-self recognition by the immune system.

One possibility is that triggering events, by altering different but specific self-components, would result in abnormal antigen processing. In certain individuals expressing a particular MHC specificity, the abnormal peptides could be recognized by non-tolerized T cells and trigger an immune response (1).

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We have previously explored the GP antigen to identify biological features of relevance in autoimmune pathogenesis. Since the NC1 domain is a highly conserved domain among species and between the different type IV collagen α chains ($\alpha 1-\alpha 6$) (2), the exclusive involvement of the human $\alpha 3(IV)NC1$ in a natural autoimmune response suggests that this domain has structural and/or biological peculiarities of pathogenic relevance. Consistent with this, the N-terminus of the human antigen is highly divergent, and it contains a unique five-residue motif (KRGDS⁹) that conforms to a functional phosphorylation site for type A protein kinases (3, 4). Furthermore, the human a3 gene, but not the other related human or homologous genes from other species, is alternatively spliced and generates multiple transcripts also containing the phosphorylatable N-terminal region (5-7). Recent studies indicate that the phosphorylation of the N-terminus of the GP antigen by cAMP-dependent protein kinase is up regulated by the presence of the alternative products (see Example 3 below). Specific serine phosphorylation and premRNA alternative splicing are also associated with the biology of other autoantigens including the acetylcholine receptor and myelin basic protein (MBP) (4). The latter is suspected to be the major antigen in multiple sclerosis (MS), another exclusively human autoimmune disease in which the immune system targets the white matter of the central nervous system. GP disease and MS are human disorders that display a strong association with the same HLA class II haplotype (HLA DRB1*1501)(32, 33). This, along with the recent report of death by GP disease of an MS patient carrying this HLA specificity (34), supports the existence of common pathogenic events in these human disorders.

Thus, specific serine/threonine phosphorylation may be a major biological difference between the human GP antigen, the GP antigens of other species, and the homologous domains from the other human $\alpha(IV)$ chains, and might be important in pathogenesis (1, 4).

Therefore, the identification and isolation of the specific serine/threonine kinase that phosphorylates the N-terminal region of the human GP antigen would be very

advantageous for the diagnosis and treatment of GP syndrome, and possibly for other autoimmune disorders.

5 Summary of the Invention

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The present invention fulfills the need in the art for the identification and isolation of a serine/threonine kinase that specifically binds to and phosphorylates the unique N-terminal region of the human GP antigen. In one aspect, the present invention provides nucleic acid sequences encoding various forms of the Goodpasture antigen binding protein (GPBP), as well as recombinant expression vectors operatively linked to the GPBP-encoding sequences.

In another aspect, the present invention provides host cells that have been transfected with the recombinant expression vectors. In a further aspect, the present invention provides substantially purified GPBP and antibodies that selectively bind to GPBP. In still further aspect, the invention provides methods for detecting the presence of GPBP or nucleic acids encoding GPBP.

In a further aspect, the present invention provides methods for detecting the presence of an autoimmune condition or apoptosis, which comprises detecting an increase in the expression of GPBP in a tissue compared to a control tissue.

In another aspect, the present invention provides methods and pharmaceutical compositions for treating an autoimmune disorder, apoptosis, or a tumor, comprising modifying the expression or activity of GPBP in a patient in need thereof.

Brief Description of the Figures

Figure 1. Nucleotide and derived amino acid sequences of n4'. The denoted structural features are from 5' to 3'end: the cDNA present in the original clone (HeLa1) (dotted box), which contains the PH homology domain (in black) and the Ser-Xaa-Yaa repeat (in gray); the heptad repeat of the predictable coiled-coil structure (open box) containing the bipartite nuclear localization signal (in gray); and a serine-rich domain (filled gray box). The asterisks denote the positions of in frame stop codons.

Figure 2. Distribution of GPBP in human tissues (Northern blot) and in eukaryotic species (Southern blot). A random primed ³²P-labeled HeLa1 cDNA probe

was used to identify homologous messages in a Northern blot of poly(A⁺)RNA from the indicated human tissues (panel A) or in a Southern blot of genomic DNA from the indicated eukaryotic species (panel B). Northern hybridization was performed under highly stringent conditions to detect perfect matching messages and at low stringency in the Southern to allow the detection of messages with mismatches. No appreciable differences in the quality and amount of each individual poly A+ RNA was observed by denaturing gel electrophoresis or when probing a representative blot from the same lot with human β -actin cDNA. The numbers denote the position and the sizes in kb of the RNA or DNA markers used.

Figure 3. Experimental determination of the translation start site. In (A), the two cDNAs present in pc-n4' and pc-FLAG-n4' plasmids used for transient expression are represented as black lines. The relative position of the corresponding predicted (n4') or engineered (FLAG-n4') translation start site is indicated (Met). In (B), the extracts from control (-), pc-n4'(n4') or pc-FLAG-n4' (FLAG-n4') transfected 293 cells were subjected to SDS-PAGE under reducing conditions in 10% gels. The separated proteins were transferred to a PVDF membrane (Millipore) and blotted with the indicated antibodies. The numbers and bars indicate the molecular mass in kDa and the relative positions of the molecular weight markers, respectively.

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Figure 4. Characterization of rGPBP from yeast and 293 cells. In (A), 1 μg (lane 1) or 100 ng (lanes 2 and 3) of yeast rGPBP were analyzed by reducing SDS-PAGE in a 10% gel. The separated proteins were stained with Coomassie blue (lane 1) or transferred and blotted with anti-FLAG antibodies (lane 2) or Mab14, a monoclonal antibody against GPBP (lane 3). In (B), the cell extracts from GPBP-expressing yeast were analyzed as in A and blotted with anti-FLAG (lane 1), anti-PSer (lane 2), anti-PThr (lane 3) or anti-PTyr (lane 4) monoclonal antibodies respectively. In (C), 200 ng of either yeast rGPBP (lane 1), dephosphorylated yeast rGPBP (lane 2) or 293 cells-derived rGPBP (lane 3) were analyzed as in B with the indicated antibodies. In (D), similar amounts of H₃³²PO₄-labeled non-transfected (lanes 1), stable pc-n4' transfected (lanes 2) or transient pc-FLAG-n4' expressing (lanes 3) 293 cells were lysed, precipitated with the indicated antibodies and analyzed by SDS-PAGE and autoradiography. The molecular weight markers are represented with numbers and bars as in Figure 3. The arrows indicate the position of the rGPBP.

Figure 5. Recombinant GPBP contains a serine/threonine kinase that specifically phosphorylates the N-terminal region of the human GP antigen. To assess phosphorylation, approximately 200 ng of yeast rGPBP was incubated with $[\gamma]^{32}$ P-ATP in the absence (A and B) or presence of GP antigen-derived material (C). In (A), the mixture was subjected to reducing SDS-PAGE (10% gel) and autoradiographed. In (B), the mixture was subjected to 32 P-phosphoamino acid analysis by two-dimensional thin-layer chromatography. The dotted circles indicate the position of ninhydrin stained phosphoamino acids. In (C), the phosphorylation mixtures of the indicated GP-derived material were analyzed by SDS-PAGE (15% gel) and autoradiography (GPpep1 and GPpep1Ala⁹) or immunoprecipitated with Mab 17, a monoclonal antibody that specifically recognize GP antigen from human and bovine origin, and analyzed by SDS-PAGE (12.5%) and autoradiography (rGP, GP). The relative positions of rGPBP (A), rGP antigen and the native human and bovine GP antigens (C) are indicated by arrows. The numbers and bars refer to molecular weight markers as in previous Figures.

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Figure 6. In-blot renaturation of the serine/threonine kinase present in rGPBP. Five micrograms of rGPBP from yeast were in-blot renatured. The recombinant material was specifically identified by anti-FLAG antibodies (lane 1) and the *in situ* ³²P-incorporation detected by autoradiography (lane 2). The numbers and bars refer to molecular weight markers as in previous Figures. The arrow indicates the position of the 89 kDa rGPBP polypeptide.

Figure 7. Immunological localization of GPBP in human tissues. Rabbit serum against the N-terminal region of GPBP (1:50) was used to localize GPBP in human tissues. The tissues shown are kidney (A) glomerulus (B), lung (C), alveolus (D), liver (E), brain (F), testis (G), adrenal gland (H), pancreas (I) and prostate (J). Similar results were obtained using anti-GPBP affinity-purified antibodies or a pool of culture medium from seven different GPBP-specific monoclonal antibodies (anti-GPBP Mabs 3, 4, 5, 6, 8, 10 and 14). Rabbit pre-immune serum did not stain any tissue structure in parallel control studies. Magnification was 40X except in B and D where it was 100X.

Figure 8. GPBPΔ26 is a splicing variant of GPBP. (A) Total RNA from normal skeletal muscle was retrotranscribed using primer 53c and subsequently

subjected to PCR with primers 11m-53c (lane 2) or 15m-62c (lane 4). Control amplifications of a plasmid containing GPBP cDNA using the same pairs of primers are shown in lanes 1 and 3. Numbers on the left and right refer to molecular weight in base pairs. The region missing in the normal muscle transcript was identified and its nucleotide sequence (lower case) and deduced amino acid sequence (upper case) are shown in (B). A clone of genomic DNA comprising the cDNA region of interest was sequenced and its structure is drawn in (C), showing the location and relative sizes of the 78-bp exon spliced out in GPBPΔ26 (black box), adjacent exons (gray boxes), and introns (lines). The size of both intron and exons is given and the nucleotide sequence of intron-exon boundaries is presented, with consensus for 5' and 3' splice sites shown in bold case.

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Figure 9. Differential expression of GPBP and GPBPΔ26. Fragments representing the 78-bp exon (GPBP) or flanking sequences common to both isoforms (GPBP/GPBPΔ26) were ³²P-labeled and used to hybridize human tissue and tumor cell line Northern blots (CLONTECH). The membranes were first hybridized with GPBP-specific probe, stripped and then reanalyzed with GPBP/GPBPΔ26 probe. Washing conditions were less stringent for GPBP-specific probe (0.1% SSPE, 37°C or 55°C) than for the GPBP/GPBPΔ26 (0.1% SSPE, 68°C) to increase GPBP and GPBPΔ26 signals respectively. No detectable signal was obtained for the GPBP probe when the washing program was at 68°C (not shown).

Figure 10. GPBPΔ26 displays lower phosphorylating activity than GPBP. (A) Recombinantly-expressed, affinity-purified GPBP (rGPBP) (lanes 1) or rGPBPΔ26 (lanes 2) were subjected to SDS-PAGE under reducing conditions and either Coomasie blue stained (2 μg per lane) or blotted (200ng per lane) with monoclonal antibodies recognizing the FLAG sequence (α-FLAG) or GPBP/GPBPΔ26 (Mab14). (B) 200 ng of rGPBP (lanes 1) or rGPBPΔ26 (lanes 2) were in vitro phosphorylated without substrate to assay auto-phosphorylation (left), or with 5 nmol GPpep1 to measure transphosphorylation activity (right). An arrowhead indicates the position of the peptide. (C) 3 μg of rGPBP (lane 1) or rGPBPΔ26 (lane 2) were in-blot renatured as described under Material and Methods. The numbers and bars indicate the molecular mass in kDa and the relative position of the molecular weight markers, respectively.

Figure 11. rGPBP and rGPBPA26 form very active high molecular weight aggregates. About 300 µg of rGPBP (A) or rGPBPA26 (B) were subjected to gel filtration HPLC as described under Material and Methods. Vertical arrowheads and numbers respectively indicate the elution profile and molecular mass (kDa) of the molecular weight standards used. Larger aggregates eluted in the void volume (I), and the bulk of the material present in the samples eluted in the fractionation range of the column as a second peak between the 669 and 158 kDa markers (II). Fifteen microliters of the indicated minute fractions were subjected to SDS-PAGE and Coomasie blue staining. Five microliters of the same fractions were in vitro phosphorylated as described in Materials and Methods, and the reaction stopped by boiling in SDS sample buffer. The fractions were loaded onto SDS-PAGE, transferred to PVDF and autoradiographed for 1 or 2 hours using Kodak X-Omat films and blotted using anti-FLAG monoclonal antibodies (Sigma).

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Figure 12. Self-interaction of GPBP and GPBP Δ 26 assessed by a yeast two-hybrid system. (A) Cell transfected for the indicated combinations of plasmids were selected on leucine-tryptophan-deficient medium (-Trp, -Leu), and independent transformants restreaked onto histidine-deficient plates (-Trp, -Leu, -His) in the presence or absence of 1 mM 3-amino-triazole (3-AT), to assess interaction. The picture was taken 3 days after streaking. (B) The bars represent mean values in β -galactosidase arbitrary units of four independent β -galactosidase in-solution assays.

Figure 13. GPBP is expressed associated with endothelial and glomerular basement membranes. Paraffin embedded sections of human muscle (A) or renal cortex (B, C) were probed with GPBP-specific antibodies (A,B) or with Mab189, a monoclonal antibody specific for the human $\alpha 3$ (IV)NC1 (C). Frozen sections of human kidney (D-F) were probed with Mab17, a monoclonal antibody specific for the $\alpha 3$ (IV)NC1 domain (D), GPBP-specific antibodies (E), or sera from a GP patient (F). Control sera (chicken pre-immune and human control) did not display tissue-binding in parallel studies (not shown).

Figure 14. GPBP is expressed in human but not in bovine and murine renal cortex. Cortex from human (A, D), bovine (B, E) or murine (C, F) kidney were paraffin

embedded and probed with either GPBP-specific antibodies (A-C) or GPBP/GPBPΔ26-specific antibodies (D-F).

Figure 15. GPBP is highly expressed in several autoimmune conditions. Skeletal muscle total RNA from a control individual (lane 1) or from a GP patient (lane 2) was subjected to RT-PCR as in Fig.8, using the oligonucleotides 15m and 62c in the amplification program. Frozen (B-D) or paraffin embedded (E-G) human control skin (B, E) or skin affected by SLE (C, F) or lichen planus (D, G) were probed with GPBP-specific antibodies.

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Figure 16. Phosphorylation of GP alternative splicing products by PKA. In left panel, equimolecular amounts of rGP (lanes 1), rGPΔV (lanes 2), rGPΔIII (lanes 3) or rGPAIII/IV/V (lanes 4), equivalent to 500 ng of the GP were phosphorylated at the indicated ATP concentrations. One-fifth of the total phosphorylation reaction mixture was separated by gel electrophoresis and transferred to PVDF, autoradiographed (shown) and the proteins blotted with M3/1, a specific monoclonal antibody recognizing all four species (shown) or using antibodies specific for each individual Cterminal region (not shown). Arrowheads indicate the position of each recombinant protein, from top to bottom, GP, GPAV and, GPAIII-GPAIII/IV/V which displayed the same mobilities. Right panel: purified a3(IV)NC1 domain or hexamer was phosphorylated with PKA and 0.1 µM ATP in the absence (lanes 1) or in the presence of 10 nmol of peptides representing the C-terminal region of either GPAIII (lanes 2) or GPAIII/IV/V (lanes 3). Where indicated the phosphorylation mixtures of purified $\alpha 3(IV)NC1$ domain were V8 digested and immunoprecipitated with antibodies specific for the N terminus of the human $\alpha 3(IV)NC1$ domain (3). Bars and numbers indicate the position and sizes (kDa) of the molecular weight markers.

Figure 17. Sequence alignment of GPΔIII and MBP. The phosphorylation sites for PKA (boxed) and the structural similarity for the sites at Ser 8 and 9 of MBP and GPΔIII respectively are shown (underlined). The identity (vertical bars) and chemical homology (dots) of the corresponding exon II (bent arrow) of both molecular species are indicated. The complete sequence of GPΔIII from the collagenase cleavage site (72-residues) is aligned with the 69-N terminal residues of MBP comprising the exon I and ten residues of the exon II.

Figure 18. Phosphorylation of rec mbinant MBP proteins by PKA. About 200 ng of rMBP (lane 1), or Ser to Ala mutants thereof in position 8 (lane 2) or 57 (lane 3), or rMPBΔII (lane 4) or Ser to Ala mutants thereof in position 8 (lane 5) or 57 (lane 6), were phosphorylated by PKA and 0.1 μM ATP. The mixtures were subjected to SDS-PAGE, transferred to PVDF and autoradiographed (Phosphorylation) and the individual molecular species blotted with monoclonal antibodies against human MBP obtained from Roche Molecular Biochemicals (Western).

Figure 19. Phosphorylation of recombinant MBP proteins by GPBP. About 200 ng of rMBP (lane 1), or Ser to Ala mutants thereof in positions 8 (lane 2) or 57 (lane 3), or rMPBAII (lane 4), or Ser to Ala mutants thereof in positions 8 (lane 5) or 57 (lane 6), were subjected to SDS-PAGE, transferred to PVDF, and the area containing the proteins visualized with Ponceau and stripped out. The immobilized proteins were in situ phosphorylated with rGPBP as described in Materials and Methods, autoradiographed (Phosphorylation) and subsequently blotted as in Fig. 18 (Western).

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Figure 20. Regulation of the GPBP by the C terminal region of GPΔIII. About 200 ng of rGPBP were in vitro phosphorylated with 150 μM ATP in the absence (lane 1) or in the presence of 5 nmol of GPΔIII-derived peptide synthesized either using Boc- (lane 2) or Fmoc- (lane 3) chemistry. The reaction mixtures were subjected to SDS-PAGE, transferred to PVDF and autoradiographed to asses autophosphorylation, and subsequently blotted with anti-FLAG monoclonal antibodies (Sigma) to determine the amount of recombinant material present (Western).

25 Detailed Description of the Invention

All references cited are herein incorporated by reference in their entirety.

The abbreviations used herein are: bp, base pair; DTT, dithiothreitol; DMEM, Dulbecco's modified Eagle's medium; EDTA, ethylenediamine tetraacetic acid; EGTA, ethylene glycol-bis(β-aminoethyl ether) N,N,N',N'-tetraacetic acid; GP, Goodpasture; rGPΔIII, rGPΔIII/IV/V and rGPΔV, recombinant material representing the alternative forms of the Goodpasture antigen resulting from splicing out exon III, exon III, IV and V or exon V, respectively; GPBP and rGPBP, native and recombinant Goodpasture

antigen binding protein; GPBPA26 and rGPBPA26, native and recombinant alternative form of the GPBP; GST, glutathione S-transferase; HLA, human lymphocyte antigens; HPLC, high performance liquid chromatography; Kb, thousand base pairs; kDa, thousand daltons; MBP, rMBP, native and recombinant 21 kDa myelin basic protein; MBPΔII and rMBPΔII, native and recombinant 18.5 kDa myelin basic protein that results from splicing out exon II; MBPAV and MBPAII/V, myelin basic protein alternative forms resulting from splicing out exon V and exons II and V, respectively; MHC, major histocompatibility complex; NC1, non-collagenous domain; PH, PMSF, PKA, cAMP-dependent protein kinase; pleckstrin homology; phenylmethylsulfonyl fluoride; SDS-PAGE, sodium dodecylsulfate polyacrylamide gel electrophoresis; TBS, tris buffered saline.

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Within this application, unless otherwise stated, the techniques utilized may be found in any of several well-known references such as: *Molecular Cloning: A Laboratory Manual* (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press), *Gene Expression Technology* (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic Press, San Diego, CA), "Guide to Protein Purification" in *Methods in Enzymology* (M.P. Deutshcer, ed., (1990) Academic Press, Inc.); *PCR Protocols: A Guide to Methods and Applications* (Innis, et al. 1990. Academic Press, San Diego, CA), *Culture of Animal Cells: A Manual of Basic Technique, 2nd Ed.* (R.I. Freshney. 1987. Liss, Inc. New York, NY), *Gene Transfer and Expression Protocols*, pp. 109-128, ed. E.J. Murray, The Humana Press Inc., Clifton, N.J.), and the Ambion 1998 Catalog (Ambion, Austin, TX).

As used herein, the term "GPBP" refers to Goodpasture binding protein, and includes both monomers and oligomers thereof. Human (SEQ ID NO:2), mouse (SEQ ID NO:4), and bovine GPBP sequences (SEQ ID NO:6) are provided herein.

As used herein, the term "GPBP Δ 26" refers to Goodpasture binding protein deleted for the 26 amino acid sequence shown in SEQ ID NO:14, and includes both monomers and oligomers thereof. Human (SEQ ID NO:8), mouse (SEQ ID NO:10), and bovine GPBP sequences (SEQ ID NO:12) are provided herein.

As used herein the term "GPBPpep1" refers to the 26 amino acid peptide shown in SEQ ID NO:14, and includes both monomers and oligomers thereof.

As used herein, the term "GP antigen" refers to the $\alpha 3$ NC1 domain of type IV collagen.

As used herein, "MBP" refers to myelin basic protein.

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In one aspect, the present invention provides isolated nucleic acids that encode GPBP, GPBPΔ26, and GPBPpep1, and mutants or fragments thereof. In one embodiment, the isolated nucleic acids comprise sequences substantially similar to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, or SEQ ID NO:25, or fragments thereof.

In another aspect, the present invention provides isolated nucleic acids that encode alternative products of the GP antigen or MBP. In one embodiment, the isolated nucleic acids comprise sequences that encode peptides substantially similar to SEQ ID NO:43 and SEQ ID NO:44.

The phrase "substantially similar" is used herein in reference to the nucleotide sequence of DNA or RNA, or the amino acid sequence of protein, having one or more conservative or non-conservative variations from the disclosed sequences, including but not limited to deletions, additions, or substitutions, wherein the resulting nucleic acid and/or amino acid sequence is functionally equivalent to the sequences disclosed herein. Functionally equivalent sequences will function in substantially the same manner to produce substantially the same protein disclosed herein. For example, functionally equivalent DNAs encode proteins that are the same as those disclosed herein or that have one or more conservative amino acid variations, such as substitution of a non-polar residue for another non-polar residue or a charged residue for a similarly charged residue. These changes include those recognized by those of skill in the art as substitutions that do not substantially alter the tertiary structure of the protein.

In practice, the term substantially similar means that DNA encoding two proteins hybridize to one another under conditions of moderate to high stringency, and encode proteins that have either the same sequence of amino acids, or have changes in sequence that do not alter their structure or function. As used herein, substantially similar sequences of nucleotides or amino acids share at least about 70% identity, more preferably at least about 80% identity, and most preferably at least about 90% identity. It is recognized, however, that proteins (and DNA or mRNA encoding such proteins)

containing less than the above-described level of homology arising as splice variants or that are modified by conservative amino acid substitutions (or substitution of degenerate codons) are contemplated to be within the scope of the present invention.

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Stringency of hybridization is used herein to refer to conditions under which nucleic acid hybrids are stable. As known to those of skill in the art, the stability of hybrids is reflected in the melting temperature (T_M) of the hybrids. T_M decreases approximately 1-1.5°C with every 1% decrease in sequence homology. In general, the stability of a hybrid is a function of sodium ion concentration and temperature. Typically, the hybridization reaction is performed under conditions of lower stringency, followed by washes of varying, but higher, stringency. Reference to hybridization stringency relates to such washing conditions. Thus, as used herein, moderate stringency refers to conditions that permit hybridization of those nucleic acid sequences that form stable hybrids in 0.1% SSPE at 37°C or 55°C, while high stringency refers to conditions that permit hybridization of those nucleic acid sequences that form stable hybrids in 0.1%SSPE at 65°C. It is understood that these conditions may be duplicated using a variety of buffers and temperatures and that they are not necessarily precise. Denhardt's solution and SSPE (see, e.g., Sambrook, Fritsch, and Maniatis, in: Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989) are well known to those of skill in the art, as are other suitable hybridization buffers.

The isolated nucleic acid sequence may comprise an RNA, a cDNA, or a genomic clone with one or more introns. The isolated sequence may further comprise additional sequences useful for promoting expression and/or purification of the encoded protein, including but not limited to polyA sequences, modified Kozak sequences, and sequences encoding epitope tags, export signals, and secretory signals, nuclear localization signals, and plasma membrane localization signals.

In another aspect, the present invention provides recombinant expression vectors comprising nucleic acid sequences that express GPBP, GPBPΔ26, or GPBPpep1, and mutants or fragments thereof. In one embodiment, the vectors comprise nucleic acid sequences that are substantially similar to the sequences shown in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, or SEQ ID NO:25, or fragments thereof.

In another aspect, the present invention provides recombinant expression vectors comprising nucleic acid sequences that express peptides that are substantially similar to the amino acid sequence shown in SEQ ID NO:43, SEQ ID NO:44, or peptide fragments thereof.

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"Recombinant expression vector" includes vectors that operatively link a nucleic acid coding region or gene to any promoter capable of effecting expression of the gene product. The promoter sequence used to drive expression of the disclosed nucleic acid sequences in a mammalian system may be constitutive (driven by any of a variety of promoters, including but not limited to, CMV, SV40, RSV, actin, EF) or inducible (driven by any of a number of inducible promoters including, but not limited to, tetracycline, ecdysone, steroid-responsive). The construction of expression vectors for use in transfecting prokaryotic cells is also well known in the art, and thus can be accomplished via standard techniques. (See, for example, Sambrook, Fritsch, and Maniatis, in: Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, 1989; Gene Transfer and Expression Protocols, pp. 109-128, ed. E.J. Murray, The Humana Press Inc., Clifton, N.J.), and the Ambion 1998 Catalog (Ambion, Austin, TX)

The expression vector must be replicable in the host organisms either as an episome or by integration into host chromosomal DNA. In a preferred embodiment, the expression vector comprises a plasmid. However, the invention is intended to include other expression vectors that serve equivalent functions, such as viral vectors.

In a further aspect, the present invention provides host cells that have been transfected with the recombinant expression vectors disclosed herein, wherein the host cells can be either prokaryotic or eukaryotic. The cells can be transiently or stably transfected. Such transfection of expression vectors into prokaryotic and eukaryotic cells can be accomplished via any technique known in the art, including but not limited to standard bacterial transformations, calcium phosphate co-precipitation, electroporation, or liposome mediated-, DEAE dextran mediated-, polycationic mediated-, or viral mediated transfection. (See, for example, *Molecular Cloning: A Laboratory Manual* (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press; Culture of Animal Cells: A Manual of Basic Technique, 2nd Ed. (R.I. Freshney, 1987. Liss, Inc. New York, NY),

In a still further aspect, the present invention provides substantially purified GPBP, GPBPΔ26, and GPBPpep1, and mutants or fragments thereof. In one embodiment, the amino acid sequence of the substantially purified protein is substantially similar to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, or peptide fragments thereof.

In another aspect, the present invention provides substantially purified alternative products of the GP antigen and MBP. In one embodiment, the amino acid sequence of the substantially purified polypeptide is substantially similar to SEQ ID NO:43, SEQ ID NO:44, or peptide fragments thereof.

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As used herein, the term "substantially purified" means that the protein has been separated from its in vivo cellular environments. Thus, the protein can either be purified from natural sources, or recombinant protein can be purified from the transfected host cells disclosed above. In a preferred embodiment, the proteins are produced by the transfected cells disclosed above, and purified using standard techniques. (See for example, *Molecular Cloning: A Laboratory Manual* (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press.)) The protein can thus be purified from prokaryotic or eukaryotic sources. In various further preferred embodiments, the protein is purified from bacterial, yeast, or mammalian cells.

The protein may comprise additional sequences useful for promoting purification of the protein, such as epitope tags and transport signals. Examples of such epitope tags include, but are not limited to FLAG (Sigma Chemical, St. Louis, MO), myc (9E10) (Invitrogen, Carlsbad, CA), 6-His (Invitrogen; Novagen, Madison, WI), and HA (Boehringer Manheim Biochemicals). Examples of such transport signals include, but are not limited to, export signals, secretory signals, nuclear localization signals, and plasma membrane localization signals.

In another aspect, the present invention provides antibodies that selectively bind to GPBP, GPBPA26, or GPBPpep1. In one aspect, the antibodies selectively bind to a protein comprising a sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, or peptide fragments thereof. Such antibodies can be produced by immunization of a host

animal with either the complete GPBP, or with antigenic peptides thereof. The antibodies can be either polyclonal or monoclonal.

In another aspect, the present invention provides antibodies that selectively bind to a polypeptide comprising an amino acid sequence substantially similar to a sequence selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO: 46, SEQ ID NO:50, SEQ ID NO:54, or antigenic fragments thereof. The antibodies can be either polyclonal or monoclonal.

Antibodies can be made by well-known methods, such as described in Harlow and Lane, Antibodies; A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., (1988). In one example, preimmune serum is collected prior to the first immunization. Substantially purified proteins of the invention, or antigenic fragments thereof, together with an appropriate adjuvant, is injected into an animal in an amount and at intervals sufficient to elicit an immune response. Animals are bled at regular intervals, preferably weekly, to determine antibody titer. The animals may or may not receive booster injections following the initial immunization. At about 7 days after each booster immunization, or about weekly after a single immunization, the animals are bled, the serum collected, and aliquots are stored at about -20° C. Polyclonal antibodies against the proteins and peptides of the invention can then be purified directly by passing serum collected from the animal through a column to which non-antigen-related proteins prepared from the same expression system without GPBP-related proteins bound.

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Monoclonal antibodies can be produced by obtaining spleen cells from the animal. (See Kohler and Milstein, Nature 256, 495-497 (1975)). In one example, monoclonal antibodies (mAb) of interest are prepared by immunizing inbred mice with the proteins or peptides of the invention, or an antigenic fragment thereof. The mice are immunized by the IP or SC route in an amount and at intervals sufficient to elicit an immune response. The mice receive an initial immunization on day 0 and are rested for about 3 to about 30 weeks. Immunized mice are given one or more booster immunizations of by the intravenous (IV) route. Lymphocytes, from antibody positive mice are obtained by removing spleens from immunized mice by standard procedures known in the art. Hybridoma cells are produced by mixing the splenic lymphocytes with an appropriate fusion partner under conditions which will allow the formation of

stable hybridomas. The antibody producing cells and fusion partner cells are fused in polyethylene glycol at concentrations from about 30% to about 50%. Fused hybridoma cells are selected by growth in hypoxanthine, thymidine and aminopterin supplemented Dulbecco's Modified Eagles Medium (DMEM) by procedures known in the art. Supernatant fluids are collected from growth positive wells and are screened for antibody production by an immunoassay such as solid phase immunoradioassay. Hybridoma cells from antibody positive wells are cloned by a technique such as the soft agar technique of MacPherson, Soft Agar Techniques, in Tissue Culture Methods and Applications, Kruse and Paterson, Eds., Academic Press, 1973.

To generate such an antibody response, the proteins of the present invention are typically formulated with a pharmaceutically acceptable carrier for parenteral administration. Such acceptable adjuvants include, but are not limited to, Freund's complete, Freund's incomplete, alum-precipitate, water in oil emulsion containing Corynebacterium parvum and tRNA. The formulation of such compositions, including the concentration of the polypeptide and the selection of the vehicle and other components, is within the skill of the art.

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The term antibody as used herein is intended to include antibody fragments thereof which are selectively reactive with the proteins and peptides of the invention, or fragments thereof. Antibodies can be fragmented using conventional techniques, and the fragments screened for utility in the same manner as described above for whole antibodies. For example, $F(ab')_2$ fragments can be generated by treating antibody with pepsin. The resulting $F(ab')_2$ fragment can be treated to reduce disulfide bridges to produce Fab' fragments.

In a further aspect, the invention provides methods for detecting the presence of the proteins or peptides of the invention in a protein sample, comprising providing a protein sample to be screened, contacting the protein sample to be screened with an antibody against the proteins or peptides of the invention, and detecting the formation of antibody-antigen complexes. The antibody can be either polyclonal or monoclonal as described above, although monoclonal antibodies are preferred. As used herein, the term "protein sample" refers to any sample that may contain the proteins or peptides of the invention, and fragments thereof, including but not limited to tissues and portions thereof, tissue sections, intact cells, cell extracts, purified or partially purified protein

samples, bodily fluids, nucleic acid expression libraries. Accordingly, this aspect of the present invention may be used to test for the presence of GPBP, GPBPΔ26, GPBPpep1, or alternative products of the GP antigen in these various protein samples by standard techniques including, but not limited to, immunolocalization, immunofluorescence analysis, Western blot analysis, ELISAs, and nucleic acid expression library screening, (See for example, Sambrook et al, 1989.) In one embodiment, the techniques may determine only the presence or absence of the protein or peptide of interest. Alternatively, the techniques may be quantitative, and provide information about the relative amount of the protein or peptide of interest in the sample. For quantitative purposes, ELISAs are preferred.

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Detection of immunocomplex formation between the proteins or peptides of the invention, or fragments thereof, and their antibodies or fragments thereof, can be For example, detection of accomplished by standard detection techniques. immunocomplexes can be accomplished by using labeled antibodies or secondary antibodies. Such methods, including the choice of label are known to those ordinarily skilled in the art. (Harlow and Lane, Supra). Alternatively, the polyclonal or monoclonal antibodies can be coupled to a detectable substance. The term "coupled" is used to mean that the detectable substance is physically linked to the antibody. Suitable detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, \u03b3-galactosidase, or acetylcholinesterase. Examples of suitable prosthetic-group complexes include streptavidin/biotin and avidin/biotin. Examples of suitable fluorescent materials include fluorescein isothiocyanate, rhodamine, fluorescein. umbelliferone. dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin. An example of a luminescent material includes luminol. Examples of suitable radioactive material include ¹²⁵I, ¹³¹I, ³⁵S or ³H.

Such methods of detection are useful for a variety of purposes, including but not limited to detecting an autoimmune condition, identifying cells targeted for or undergoing apoptosis, immunolocalization of the proteins of interest in a tissue sample, Western blot analysis, and screening of expression libraries to find related proteins.

In yet another aspect, the invention provides methods for detecting the presence in a sample of nucleic acid sequences encoding the GPBP, GPBPA26, GPBPpep1, or alternative products of the GP antigen comprising providing a nucleic acid sample to be screened, contacting the sample with a nucleic acid probe derived from the isolated nucleic acid sequences of the invention, or fragments thereof, and detecting complex formation.

As used herein, the term "sample" refers to any sample that may contain GPBPrelated nucleic acid, including but not limited to tissues and portions thereof, tissue sections, intact cells, cell extracts, purified or partially purified nucleic acid samples, DNA libraries, and bodily fluids. Accordingly, this aspect of the present invention may be used to test for the presence of GPBP mRNA or DNA in these various samples by standard techniques including, but not limited to, in situ hybridization, Northern blotting, Southern blotting, DNA library screening, polymerase chain reaction (PCR) or reverse transcription-PCR (RT-PCR). (See for example, Sambrook et al, 1989.) In one embodiment, the techniques may determine only the presence or absence of the nucleic acid of interest. Alternatively, the techniques may be quantitative, and provide information about the relative amount of the nucleic acid of interest in the sample. For quantitative purposes, quantitative PCR and RT-PCR are preferred. Thus, in one example, RNA is isolated from a sample, and contacted with an oligonucleotide derived from the nucleic acid sequence of interest, together with reverse transcriptase under suitable buffer and temperature conditions to produce cDNAs from the GPBP-related RNA. The cDNA is then subjected to PCR using primer pairs derived from the nucleic acid sequence of interest. In a preferred embodiment, the primers are designed to detect the presence of the RNA expression product of SEQ ID NO:5, and the amount of GPBP gene expression in the sample is compared to the level in a control sample.

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For detecting the nucleic acid sequence of interest, standard labeling techniques can be used to label the probe, the nucleic acid of interest, or the complex between the probe and the nucleic acid of interest, including, but not limited to radio-, enzyme-, chemiluminescent-, or avidin or biotin-labeling techniques, all of which are well known in the art. (See, for example, *Molecular Cloning: A Laboratory Manual* (Sambrook, et al., 1989, Cold Spring Harbor Laboratory Press), *Gene Expression Technology* (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic Press, San

Diego, CA); PCR Protocols: A Guide to Methods and Applications (Innis, et al. 1990. Academic Press, San Diego, CA)).

Such methods of nucleic acid detection are useful for a variety of purposes, including but not limited to diagnosing an autoimmune condition, identifying cells targeted for or undergoing apoptosis, in situ hybridization, Northern and Southern blot analysis, and DNA library screening.

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As demonstrated in the following examples, GPBP shows preferential expression in tissue structures that are commonly targeted in naturally-occurring automimmune responses, and is highly expressed in several autoimmune conditions, including but not limited to Goodpasture Syndrome (GP), systemic lupus erythematosus (SLE), and lichen planus. Furthermore, following a similar experimental approach to that described below, recombinant proteins representing autoantigens in GP disease (α 3 Type IV collagen), SLE (P1 ribosomal phosphoprotein and Sm-D1 small nuclear ribonucleoproteins) and dermatomyositis (hystididyl-tRNA synthetase) were shown to be in vitro substrates of GPBP.

Thus, in a preferred embodiment, detection of GPBP expression is used to detect an autoimmune condition. A sample that is being tested is compared to a control sample for the expression of GPBP, wherein an increased level of GPBP expression indicates the presence of an autoimmune condition. In this embodiment, it is preferable to use antibodies that selectively bind to GPBPpep1, which is present in GPBP but not in GPBPA26.

Furthermore, as shown in the accompanying examples, GPBP is down-regulated in tumor cell lines, and the data suggest that GPBP/GPBPΔ26 are likely to be involved in cell signaling pathways that induce apoptosis, which may be up-regulated during autoimmune pathogenesis and down-regulated during cell transformation to prevent autoimmune attack to transformed cells during tumor growth. Thus, the detection methods disclosed herein can be used to detect cells that are targeted for, or are undergoing apoptosis.

In another aspect, the present invention provides a method for treating an autoimmune disorder, a tumor, or for preventing cell apoptosis comprising modification of the expression or activity of GPBP, GPBP Δ 26, or a protein comprising a polypeptide substantially similarly to GPBPpep1 in a patient in need thereof. Modifying the

expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1 can be accomplished by using specific inducers or inhibitors of GPBP expression or activity, GPBP antibodies, gene or protein therapy using GP or myelin basic protein alternative products, cell therapy using host cells expressing GP or myelin basic protein alternative products, antisense therapy, or other techniques known in the art. In a preferred embodiment, the method further comprises administering a substantially purified alternative product of the GP antigen or MBP to modify the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1. As used herein, "modification of expression or activity" refers to modifying expression or activity of either the RNA or protein product.

In a further aspect, the present invention provides pharmaceutical compositions, comprising an amount effective of substantially purified alternative products of the GP antigen or MBP to modify the expression or activity of GPBP RNA or protein, and a pharmaceutically acceptable carrier.

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For administration, the active agent is ordinarily combined with one or more adjuvants appropriate for the indicated route of administration. The compounds may be mixed with lactose, sucrose, starch powder, cellulose esters of alkanoic acids, stearic acid, talc, magnesium stearate, magnesium oxide, sodium and calcium salts of phosphoric and sulphuric acids, acacia, gelatin, sodium alginate, polyvinylpyrrolidine, and/or polyvinyl alcohol, and tableted or encapsulated for conventional administration. Alternatively, the compounds of this invention may be dissolved in saline, water, polyethylene glycol, propylene glycol, carboxymethyl cellulose colloidal solutions, ethanol, corn oil, peanut oil, cottonseed oil, sesame oil, tragacanth gum, and/or various buffers. Other adjuvants and modes of administration are well known in the pharmaceutical art. The carrier or diluent may include time delay material, such as glyceryl monostearate or glyceryl distearate alone or with a wax, or other materials well known in the art.

The present invention may be better understood with reference to the accompanying examples that are intended for purposes of illustration only and should not be construed to limit the scope of the invention, as defined by the claims appended hereto.

Example 1: Characterizati n of GPBP

Here we report the cloning and characterization of a novel type of serine/threonine kinase that specifically binds to and phosphorylates the unique N-terminal region of the human GP antigen.

MATERIALS AND METHODS

Synthetic polymers-Peptides. GPpep1, KGKRGDSGSPATWTTRGFVFT (SEQ ID NO:26), representing residues 3-23 of the human GP antigen and GPpep1Ala⁹, KGKRGDAGSPATWTTRGFVFT (SEQ ID NO:27), a mutant Ser⁹ to Ala⁹ thereof, were synthesized by MedProbe and CHIRON. FLAG peptide, was from Sigma.

Oligonucleotides. The following as well as several other GPBP-specific oligonucleotides were synthesized by Genosys and GIBCO BRL:

15 ON-GPBP-54m:

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TCGAATTCACCATGGCCCACTAGCCGACTACAAGGACGACGATG ACAAG (SEQ ID NO: 28).

ON-GPBP-55c:

CCGAGCCCGACGAGTTCCAGCTCTGATTATCCGACATCTTGTCATCG

20 TCG (SEQ ID NO:29).

ON-HNC-B-N-14m: CGGGATCCGCTAGCTAAGCCAGGCAAGGATGG (SEQ ID NO:30).

ON-HNC-B-N-16c: CGGGATCCATGCATAAATAGCAGTTCTGCTGT (SEQ ID NO:31).

Isolation and characterization of cDNA clones encoding human GPBP-Several human λ-gt11 cDNA expression libraries (cye, fetal and adult lung, kidney and HeLa S3, from CLONTECH) were probed for cDNAs encoding proteins interacting with GPpep1. Nitrocellulose filters (Millipore) prepared following standard immunoscreening procedures were blocked and incubated with 1-10 nmoles per ml of GPpep1 at 37°C. Specifically bound GPpep1 was detected using M3/1A monoclonal antibodies (7). A single clone was identified in the HeLa-derived library (HeLa1). Specificity of fusion protein binding was confirmed by similar binding to recombinant

eukaryotic human GP antigen. The EcoRI cDNA insert of HeLa1 (0.5-kb) was used to further screen the same library and to isolate overlapping cDNAs. The largest cDNA (2.4-kb) containing the entire cDNA of HeLa1 (n4') was fully sequenced.

Northern and Southern blots-Pre-made Northern and Southern blots (CLONTECH) were probed with HeLa1 cDNA following manufacturer instructions.

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Plasmid construction, expression and purification of recombinant proteins-GPBP-derived material. The original λ-gtl1 HeLa1 clone was expressed as a lysogen in E. Coli Y1089 (8). The corresponding β-galactosidase-derived fusion protein containing the N-terminal 150 residues of GPBP was purified from the cell lysate using an APTG-agarose column (Boehringer). The EcoRI 2.4-kb fragment of n4' was subcloned in Bluescribe M13+ vector (Stratagene) (BS-n4'), amplified and used for subsequent cloning. A DNA fragment containing (from 5' to 3'), an EcoRI restriction site, a standard Kozak consensus for translation initiation, a region coding for a tag peptide sequence (FLAG, DYKDDDDK (SEQ ID NO:32)), and the sequence coding for the first eleven residues of GPBP including the predicted Met, and a Ban II restriction site, was obtained by hybridizing ON-GPBP-54m and ON-GPBP-55c, and extending with modified T₇ DNA polymerase (Amersham). The resulting DNA product was digested with EcoRI and BanII, and ligated with the BanII/EcoRI cDNA fragment of BS-n4' in the EcoRI site of pHIL-D2 (Invitrogen) to produce pHIL-FLAG-n4'. This plasmid was used to obtain Muts transformants of the GS115 strain of Pichia pastoris and to express FLAG-tagged recombinant GPBP (rGPBP) either by conventional liquid culture or by fermentation procedures (Pichia Expression Kit, Invitrogen). The cell lysates were loaded onto an anti-FLAG M2 column (Sigma), the unbound material washed out with Tris buffered saline (TBS, 50 mM Tris-HCl, pH 7.4, 150 mM NaCl) or salt-supplemented TBS (up to 2M NaCl), and the recombinant material eluted with FLAG peptide. For expression in cultured human kidney-derived 293 cells (ATCC 1573-CRL), the 2.4- or 2.0-kb EcoRI cDNA insert of either BS-n4' or pHIL-FLAG-n4' was subcloned in pcDNA3 (Invitrogen) to produce pc-n4' and pc-FLAG-n4' respectively. When used for transient expression, 18 hours after transfection the cells were lysed with 3.5-4 µl/cm² of chilled lysis buffer (1% Nonidet P-40 or Triton-X100, 5mM EDTA and 1 mM PMSF in TBS) with or without 0.1% SDS, depending on whether the lysate was to be used for SDS-PAGE or FLAG-purification, respectively. For FLAG purification, the lysate of four to six 175 cm² culture dishes was diluted up to 50 ml with lysis buffer and purified as above. For stable expression, the cells were similarly transfected with pc-n4' and selected for three weeks with 800 μg/ml of G418. For bacterial recombinant expression, the 2.0-kb EcoRI cDNA fragment of pHIL-FLAG-n4' was cloned in-frame downstream of the glutathione S-transferase (GST)-encoding cDNA of pGEX-5x-1 (Pharmacia). The resulting construct was used to express GST-GPBP fusion protein in DH5α cells (9).

GP antigen-derived material. Human recombinant GP antigen (rGP) was produced in 293 cells using the pRc/CMV-BM40 expression vector containing the α 3-specific cDNA between ON-HNC-B-N-14m and ON-HNC-B-N-16c. The expression vector is a pRc/CMV (Invitrogen)-derived vector provided by Billy G. Hudson (Kansas University Medical Center) that contains cDNA encoding an initiation Met, a BM40 signal peptide followed by a tag peptide sequence (FLAG), and a polylinker cloning site. To obtain α 3-specific cDNA, a polymerase chain reaction was performed using the oligonucleotides above and a plasmid containing the previously reported α 3(IV) cDNA sequence (3) as template (clone C2). For stable expression of rGP, 293 cells were transfected with the resulting construct (α 3VLC) and selected with 400 α 9/ml of G418. The harvested rGP was purified using an anti-FLAG M2 column.

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All the constructs were verified by restriction mapping and nucleotide sequencing.

Cell culture and DNA transfection-Human 293 cells were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal calf serum. Transfections were performed using the calcium phosphate precipitation method of the Profection Mammalian Transfection Systems (Promega). Stably transfected cells were selected by their resistance to G418. Foci of surviving cells were isolated, cloned and amplified.

Antibody production-Polyclonal antibodies against the N-terminal region of GPBP. Cells expressing HeLa1 λ -gt11 as a lysogen were lysed by sonication in the presence of Laemmli sample buffer and subjected to electrophoresis in a 7.5% acrylamide preparative gel. The gel was stained with Coomassie blue and the band containing the fusion protein of interest excised and used for rabbit immunization (10). The anti-serum was tested for reactivity using APTG-affinity purified antigen. To

obtain affinity-purified antibodies, the anti-serum was diluted 1:5 with TBS and loaded onto a Sepharose 4B column containing covalently bound affinity purified antigen. The bound material was eluted and, unless otherwise indicated, used in the immunochemical studies.

Monoclonal antibodies against GPBP. Monoclonal antibodies were produced essentially as previously reported (7) using GST-GPBP. The supernatants of individual clones were analyzed for antibodies against rGPBP.

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In vitro phosphorylation assays-About 200 ng of rGPBP were incubated overnight at 30°C in 25 mM β -glycerolphosphate (pH 7.0), 0.5 mM EDTA, 0.5 mM EGTA, 8 mM MgCl₂, 5 mM MnCl₂, 1 mM DTT and 0.132 μ M γ -³²P-ATP, in the presence or absence of 0.5-1 μ g of protein substrates or 10 nmoles of synthetic peptides, in a total volume of 50 μ l.

In vivo phosphorylation assays-Individual wells of a 24-well dish were seeded with normal or with stably pc-n4' transfected 293 cells. When the cells were grown to the desired density, a number of wells of the normal 293 cells were transfected with pc-FLAG-n4'. After 12 hours, the culture medium was removed, 20 μ Ci/well of H₃³²PO4 in 100 μ l of phosphate-free DMEM added, and incubation continued for 4 hours. The cells were lysed with 300 μ l/well of TBS containing 1% Triton X-100, 2 mM EDTA, 1 mM PMSF, 50 mM NaF and 0.2 mM vanadate, and extracted with specific antibodies and Protein A-Sepharose. When anti-GPBP serum was used, the lysate was pre-cleared using pre-immune serum and Protein A-Sepharose.

In vitro dephosphorylation of rGPBP-About 1 μ g of rGPBP was dephosphorylated in 100 μ l of 10 mM Tris-acetate (pH 7.5), 10 mM magnesium acetate and 50 mM potassium acetate with 0.85 U of calf intestine alkaline phosphatase (Pharmacia) for 30 min at 30°C.

Renaturation assays-In-blot renaturation assays were performed using 1-5 μ g of rGPBP as previously described (11).

Nucleotide sequence analysis- cDNA sequence analyses were performed by the dideoxy chain termination method using $[\alpha]^{35}$ S-dATP, modified T_7 DNA polymerase (Amersham) and universal or GPBP-specific primers (8-10).

³²P-Phosphoamino acid analysis-Immunopurified rGPBP or HPLC gelfiltration fractions thereof containing the material of interest were phosphorylated, hydrolyzed and analyzed in one dimensional (4) or two dimensional thin layer chromatography (12). When performing two dimensional analysis, the buffer for the first dimension was formic acid:acetic acid:water (1:3.1:35.9) (pH 1.9) and the buffer for the second dimension was acetic acid:pyridine:water (2:0.2:37.8) (pH 3.5). Amino acids were revealed with ninhydrin, and ³²P-phosphoamino acids by autoradiography.

Physical methods and immunochemical techniques-SDS-PAGE and Western-blotting were performed as in (4). Immunohistochemistry studies were done on human multi-tissue control slides (Biomeda, Biogenex) using the ABC peroxidase method (13).

Computer analysis-Homology searches were carried out against the GenBank and SwissProt databases with the BLAST 2.0 (14) at the NCBI server, and against the TIGR Human Gene Index database for expressed sequence tags, using the Institute for Genomic Research server. The search for functional patterns and profiles was performed against the PROSITE database using the ProfileScan program at the Swiss Institute of Bioinformatics (15). Prediction of coiled-coil structures was done at the Swiss Institute for Experimental Cancer Research using the program Coils (16) with both 21 and 28 residue windows.

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RESULTS

Molecular cloning of GPBP-To search for proteins specifically interacting with the divergent N-terminal region of the human GP antigen, a 21-residue peptide (GPpep1; SEQ ID NO:26)), encompassing this region and flanking sequences, and specific monoclonal antibodies against it were combined to screen several human cDNA expression libraries. More than 5 x 10⁶ phages were screened to identify a single HeLaderived recombinant encoding a fusion protein specifically interacting with GPpep1 without disturbing antibody binding.

Using the cDNA insert of the original clone (HeLa1), we isolated a 2.4-kb cDNA (n4') that contains 408-bp of 5'-untranslated sequence, an open reading frame (ORF) of

1872-bp encoding 624 residues, and 109-bp of 3'-untranslated sequence (Fig. 1) (SEQ ID NO:1-2). Other structural features are of interest. First, the predicted polypeptide (hereinafter referred to as GPBP) has a large number of phosphorylatable (17.9%) and acidic (16%) residues unequally distributed along the sequence. Serine, which is the most abundant residue (9.3%), shows preference for two short regions of the protein, where it comprises nearly 40% of the amino acids, compared to an average of less than 7% throughout the rest of the polypeptide chain. It is also noteworthy that the more Nterminal, serine-rich region consists mainly of a Ser-Xaa-Yaa repeat. Acidic residues are preferentially located at the N-terminal three-quarters of the polypeptide, with nearly 18% of the residues being acidic. These residues represent only 9% in the most C-terminal quarter of the polypeptide, resulting in a polypeptide chain with two electrically opposite domains. At the N-terminus, the polypeptide contains a pleckstrin homology (PH) domain, which has been implicated in the recruitment of many signaling proteins to the cell membrane where they exert their biological activities (17). Finally, a bipartite nuclear targeting sequence (18) exists as an integral part of a heptad repeat region that meets all the structural requirements to form a coiled-coil (16).

Protein data bank searches revealed homologies almost exclusively within the approximately 100 residues at the N-terminal region harboring the PH domain. The PH domain of the oxysterol-binding protein is the most similar, with an overall identity of 33.5% and a similarity of 65.2% with GPBP. In addition, the *Caenorhabditis elegans* cosmid F25H2 (accession number Q93569) contains a hypothetical ORF that displays an overall identity of 26.5% and a similarity of 61% throughout the entire protein sequence, indicating that similar proteins are present in lower invertebrates. Several human expressed sequence tags (accession numbers AA287878, AA287561, AA307431, AA331618, AA040134, AA158618, AA040087, AA122226, AA158617, AA121104, AA412432, AA412433, AA282679 and N27578) possess a high degree of nucleotide identity (above 98%) with the corresponding stretches of the GPBP cDNA, suggesting that they represent human GPBP. Interestingly, the AA287878 EST shows a gap of 67 nucleotides within the sequence corresponding to the GPBP 5'-untranslated region, suggesting that the GPBP pre-mRNA is alternatively spliced in human tissues (not shown).

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The distribution and expression of the GPBP gene in human tissues was first assessed by Northern blot analysis (Fig. 2, panel A). The gene is expressed as two major mRNAs species between 4.4-kb and 7.5-kb in length and other minor species of shorter lengths. The structural relationship between these multiple mRNA species is not known and their relative expression varies between tissues. The highest expression level is seen in striated muscle (skeletal and heart), while lung and liver show the lowest expression levels.

Southern blot studies analysis of genomic DNA from different species indicated that homologous genes exist throughout phylogeny (Fig. 2, panel B). Consistent with the human origin of the probe, the hybridization intensities decreased in a progressive fashion as the origin of the genomic DNA moves away from humans in evolution.

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Experimental determination of the translation start site-To experimentally confirm the predicted ORF, eukaryotic expression vectors containing either the 2.4-kb of cDNA of n4', or only the predicted ORF tagged with a FLAG sequence (Fig. 3A), were used for transient expression assays in 293 cells. The corresponding extracts were analyzed by immunoblot using GPBP- or FLAG-specific antibodies. The GPBP-specific antibodies bind to a similar major polypeptide in both transfected cells, but only the polypeptide produced by the engineered construct expressed the FLAG sequence (Fig. 3B). This located the translation start site of the n4' cDNA at the predicted Met and confirmed the proposed primary structure. Furthermore, the recombinant polypeptides displayed a molecular mass higher than expected (80 versus 71 kDa) suggesting that GPBP undergoes post-translational modifications.

Expression and characterization of yeast rGPBP-Yeast expression and FLAG-based affinity-purification were combined to produce rGPBP (Fig. 4A). A major polypeptide of ~89 kDa, along with multiple related products displaying lower M_r , were obtained. The recombinant material was recognized by both anti-FLAG and GPBP-specific antibodies, guaranteeing the fidelity of the expression system. Again, however, the M_r displayed by the major product was notably higher than predicted and even higher than the M_r of the 293 cell-derived recombinant material, supporting the idea that GPBP undergoes important and differential post-translational modifications. Since phosphorylatable residues are abundant in the polypeptide chain, we investigated the existence of phosphoamino acids in the recombinant materials. By using monoclonal or

polyclonal (not shown) antibodies against phosphoserine (Pser), phosphothreonine (PThr) and phosphotyrosine (PTyr), we identified the presence of all three phosphoresidues either in yeast rGPBP (Fig. 4B) or in 293 cell-derived material (not shown). The specificity of the antibodies was further assessed by partially inhibiting their binding by the addition of 5-10 mM of the corresponding phosphoamino acid (not shown). This suggests that the phosphoresidue content varies depending upon the cell expression system, and that the M_r differences are mainly due to phosphorylation. Dephosphorylated yeast-derived material consistently displayed similar M_r to the material derived from 293 cells, and phosphoamino acid content correlates with SDS-PAGE mobilities (Fig. 4C). As an *in vivo* measurement, the phosphorylation of rGPBP in the 293 cells was assessed (Fig. 4D). Control cells (lanes 1) and cells expressing rGPBP in a stable (lanes 2) or transient (lanes 3) mode were cultured in the presence of H_3^{32} PO₄. Immunoprecipitated recombinant material contained 32 P, indicating that phosphorylation of GPBP occurred *in vivo* and therefore is likely to be a physiological process.

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The rGPBP is a serine/threonine kinase that phosphorylates the N-terminal region of the human GP antigen-Although GPBP does not contain the conserved structural regions required to define the classic catalytic domain for a protein kinase, the recent identification and characterization of novel non-conventional protein kinases (19-27) encouraged the investigation of its phosphorylating activity. Addition of $[\gamma^{32}P]ATP$ to rGPBP (either from yeast or 293 cells (not shown)) in the presence of Mn2+ and Mg2+ resulted in the incorporation of ³²P as PSer and PThr in the major and related products recognized by both anti-FLAG and specific antibodies (Fig. 5A and B), indicating that the affinity-purified material contains a Ser/Thr protein kinase. To further characterize this activity, GPpep1, GPpep1Ala9 (a GPpep1 mutant with Ser9 replaced by Ala), native and recombinant human GP antigens, and native bovine GP antigen were assayed (Fig. 5C). Affinity-purified rGPBP phosphorylates all human-derived material to a different extent. However, in similar conditions, no appreciable 32P-incorporation was observed in the bovine-derived substrate. The lower 32P incorporation displayed by GPpep1Ala9 when compared with GPpep1, and the lack of phosphorylation of the bovine antigen, indicates that the kinase present in rGPBP discriminates between human and bovine antigens, and that Ser⁹ is a target for the kinase.

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Although the purification system provides high quality material, the presence of contaminants with a protein kinase activity could not be ruled out. The existence of contaminants was also suggested by the presence of a FLAG-containing 40 kDa polypeptide, which displayed no reactivity with specific antibodies nor incorporation of ³²P in the phosphorylation assays (Fig. 4A and 5A). To precisely identify the polypeptide harboring the protein kinase activity, we performed in vitro kinase renaturation assays after SDS-PAGE and Western-blotted (Fig. 6). We successfully combined the use of specific antibodies (lane 1) and autoradiographic detection of in situ ³²P-incorporation (lane 2), and identified the 89 kDa rGPBP material as the primary polypeptide harboring the Ser/Thr kinase activity. The lack of ³²P-incorporation in the rGPBP-derived products, as well as in the 40 kDa contaminant, further supports the specificity of the renaturation assays and locates the kinase activity to the 89 kDa polypeptide. Recently, it has been shown that traces of protein kinases intimately associated with a polypeptide can be released from the blot membrane, bind to, and phosphorylate the polypeptide during the labeling step (28). To assess this possibility in our system, we performed renaturation studies using a small piece of membrane containing the 89 kDa polypeptide, either alone or together with membrane pieces representing the different regions of the blot lane. We observed similar ³²P-incorporation at the 89 kDa polypeptide regardless of the coincubated pieces (not shown), indicating that if there are co-purified protein kinases in our sample they are not phosphorylating the 89 kDa polypeptide in the renaturation assays unless they co-migrate. Co-migration does not appear to be a concern, however, since rGPBP deletion mutants (GPBPA26 and R3; see below) displaying different mobilities also have kinase activities and could be similarly in-blot renatured (not shown).

Immunohistochemical localization of the novel kinase-To investigate GPBP expression in human tissues we performed immunohistochemical studies using specific polyclonal (Fig.7) or monoclonal antibodies (not shown). Although GPBP is widely expressed in human tissues, it shows tissue and cell-specificity. In kidney, the major expression is found at the tubule epithelial cells and the glomerular mesangial cells and podocytes. At the lung alveolus, the antibodies display a linear pattern suggestive of a basement membrane localization, along with staining of pneumocytes. Liver shows low expression in the parenchyma, but high expression in biliary ducts. Expression in the central nervous system is observed in the white matter, but not in the neurons of the brain.

In testis, a high expression in the spermatogonium contrasts with the lack of expression in Sertoli cells. The adrenal gland shows a higher level of expression in cortical cells versus the medullar. In the pancreas, GPBP is preferentially expressed in Langerhans islets versus the exocrine moiety. In prostate, GPBP is expressed in the epithelial cells but not in the stroma (Fig. 7). Other locations with high expression of GPBP are striated muscle, epithelial cells of intestinal tract, and Purkinje cells of the cerebellum (not shown). In general, in tissues where GPBP is highly expressed the staining pattern is mainly diffuse cytosolic. However in certain locations there is, in addition, an important staining reinforcement at the nucleus (spermatogonium), at the plasma membrane (pneumocyte, hepatocyte, prostate epithelial cells, white matter) or at the extracellular matrix (alveolus) (Fig. 7).

DISCUSSION

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Our data show that GPBP is a novel, non-conventional serine/threonine kinase. We also present evidence that GPBP discriminates between human and bovine GP antigens, and targets the phosphorylatable region of human GP antigen in vitro. Several lines of evidence indicate that the 89 kDa polypeptide is the only kinase in the affinity purified rGPBP. First, we found no differences in auto- or trans-phosphorylation among rGPBP samples purified in the presence of 150 mM, 0.5 M, 1 M or 2 M salt (not shown), suggesting that rGPBP does not carry intimately bound kinases. Second, there is no FLAG-containing, yeast-derived kinase in our samples, since material purified using GPBP-specific antibodies shows no differences in phosphorylation (not shown). Third, a deletion mutant (GPBPA26; see below) displays reduced auto- and trans-phosphorylation activities (not shown), demonstrating that the 89 kD polypeptide is the only portion of the rGPBP with the ability to carry out phosphate transfer.

Although GPBP is not homologous to other non-conventional kinases, they share some structural features including an N-terminal α -helix coiled-coil (26, 27), serine-rich motifs (24), high phosphoamino acids content (27), bipartite nuclear localization signal (27), and the absence of a typical nucleotide or ATP binding motif (24, 27).

Immunohistochemistry studies show that GPBP is a cytosolic polypeptide also found in the nucleus, associated with the plasma membrane and likely at the extracellular matrix associated with the basement membrane, indicating that it contains the structural

requirements to reach all these destinations. The nuclear localization signal and the PH domain confer to it the potential to reach the nucleus and the cell membrane, respectively (17, 29, 30). Although GPBP does not contain the structural requirements to be exported, the 5'-end untranslated region of its mRNA includes an upstream ORF of 130 residues with an in-frame stop codon at the beginning (Fig. 1). A mRNA editing process inserting a single base pair (U) would generate an operative in-frame start site and an ORF of 754-residues containing an export signal immediately downstream of the edited Met (not shown). Polyclonal antibodies against a synthetic peptide representing part of this hypothetical extra-sequence (PRSARCQARRRRGGRTSS (SEQ ID NO:33)) display a linear vascular reactivity in human tissues suggestive of an extracellular basement membrane localization (data not shown).

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Alternatively, a splicing phenomenon could generate transcripts with additional unidentified exon(s) that would provide the structural requirements for exportation. The multiple cellular localization, the high content in PTyr, and the lack of tyrosine kinase activity in vitro, suggest that GPBP is itself the target of specific tyrosine kinase(s) and therefore likely involved in specific signaling cascade(s).

As discussed above, specific serine phosphorylation, as well as pre-mRNA alternative splicing, are associated with the biology of several autoantigens, including the GP antigen, acetylcholine receptor and myelin basic protein (MBP) (4). The latter is suspected to be the major antigen in multiple sclerosis (MS), another exclusively human autoimmune disease in which the immune system targets the white matter of the central nervous system. GP disease and MS are human disorders that display a strong association with the same HLA class II haplotype (HLA DRB1*1501)(32, 33). This, along with the recent report of death by GP disease of a MS patient carrying this HLA specificity (34), supports the existence of common pathogenic events in these human disorders.

Phosphorylation of specific serines has been shown to change intracellular proteolysis (35-40). Conceivably, alterations in protein phosphorylation can affect processing and peptide presentation, and thus mediate autoimmunity. GP antigen-derived peptide presentation by the HLA-DR15 depends more on processing than on preferences of relatively indiscriminate DR15 molecules (41), suggesting that if processing is influenced by abnormal phosphorylation, the resulting peptides would likely be presented by this HLA. Our more recent data indicate that in both the GP and MBP systems, the

production of alternative splicing products serves to regulate the phosphorylation of specific and structurally homologous PKA sites, suggesting that this or a closely related kinase is the *in vivo* phosphorylating enzyme. Alterations in the degree of antigen phosphorylation, caused either by an imbalance in alternative products, or by the action of an intruding kinase that deregulates phosphorylation of the same motifs, could lead to an autoimmune response in predisposed individuals. rGPBP phosphorylates the human GP antigen at a major PKA phosphorylation site in an apparently unregulated fashion, since the presence of specific alternative products of the GP antigen did not affect phosphorylation of the primary antigen by GPBP (not shown).

Although GPBP is ubiquitously expressed, in certain organs and tissues it shows a preference for cells and tissue structures that are target of common autoimmune responses: the Langerhans cells (type I diabetes); the white matter of the central nervous system (multiple sclerosis); the biliary ducts (primary biliary cirrhosis); the cortical cells of the adrenal gland (Addison disease); striated muscle cells (myasthenia gravis); spermatogonium (male infertility); Purkinje cells of the cerebellum (paraneoplasic cerebellar degeneration syndrome); and intestinal epithelial cells (pernicious anemia, autoimmune gastritis and enteritis). All the above observations point to this novel kinase as an attractive candidate to be considered when envisioning a model for human autoimmune disease.

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Example 2: GPBP Alternative Splicing

Here we report the existence of two isoforms of GPBP that are generated by alternative splicing of a 78-base pair (bp) long exon that encodes a 26-residue serinerich motif. Both isoforms, GPBP and GPBP Δ 26, exist as high molecular aggregates that result from polypeptide self-aggregation. The presence of the 26-residue peptide in the polypeptide chain results in a molecular species that self-interacts more efficiently and forms aggregates with higher specific activity. Finally, we present evidences supporting the observation that GPBP is implicated in human autoimmune pathogenesis.

10 MATERIAL AND METHODS.

Synthetic polymers:

Peptides. GPpep1, KGKRGDSGSPATWTTRGFVFT (SEQ ID NO:26), is described in Example 1. GPBPpep1, PYSRSSSMSSIDLVSASDDVHRFSSQ (SEQ ID NO:14), representing residues 371-396 of GPBP was synthesized by Genosys.

- Oligonucleotides. The following oligonucleotides were synthesized by Life Technologies, Inc., 5' to 3': ON-GPBP-11m, G CGG GAC TCA GCG GCC GGA TTT TCT (SEQ ID NO:34); ON-GPBP-15m, AC AGC TGG CAG AAG AGA C (SEQ ID NO:35); ON-GPBP-20c, C ATG GGT AGC TTT TAA AG (SEQ ID NO; 36); ON-GPBP-22m, TA GAA GAA CAG TCA CAG AGT GAA AAG G (SEQ ID NO:37); ON-GPBP-53c, GAATTC GAA CAA AAT AGG CTT TC (SEQ ID NO:38); ON-GPBP-56m, CCC TAT AGT CGC TCT TC (SEQ ID NO:39); ON-GPBP-57c, CTG GGA GCT GAA TCT GT (SEQ ID NO:40); ON-GPBP-62c, GTG GTT CTG CAC CAT CTC TTC AAC (SEQ ID NO:41); ON-GPBP-Δ26, CA CAT AGA TTT GTC CAA AAG GTT GAA GAG ATG GTG CAG AAC (SEQ ID NO:42).
- Reverse transcriptase and polymerase chain rection (RT-PCR). Total RNA was prepared from different control and GP tissues as described in (15). Five micrograms of total RNA was retrotranscribed using Ready-To-Go You-Prime First-Strand beads (Amersham Pharmacia Biotech) and 40 pmol of ON-GPBP-53c. The corresponding cDNA was subjected to PCR using the pairs of primers ON-GPBP-11m/ON-GPBP-53c or ON-GPBP-15m/ON-GPBP-62c. The identity of the products obtained with 15m-62c

was further confirmed by Alu I restriction. To specifically amplify GPBP transcripts, PCR was performed using primers ON-GPBP-15m/ON-GPBP-57c.

Northern hybridization studies. Pre-made human multiple-tissue and tumor cell-line Northern Blots (CLONTECH) were probed with a cDNA containing the 78-bp exon present only in GPBP or with a cDNA representing both isoforms. The corresponding cDNAs were obtained by PCR using the pair of primers ON-GPBP-56m and ON-GPBP-57c using GPBP as a template, or with primers ON-GPBP-22m and ON-GPBP-20c, using GPBPΔ26 as a template. The resulting products were random-labeled and hybridized following the manufacturers' instructions.

Plasmid construction, expression and purification of recombinant proteins. The plasmid pHIL-FLAG-n4', used for recombinant expression of FLAG-tagged GPBP in *Pichia pastoris* has been described elsewhere (4). The sequence coding for the 78-bp exon was deleted by site-directed mutagenesis using ON-GPBP-Δ26 to generate the plasmid pHIL-FLAG-n4'Δ26. Expression and affinity-purification of recombinant GPBP and GPBPΔ26 was done as in (4).

Gel-filtration HPLC. Samples of 250 µl were injected into a gel filtration PE-TSK-G4000SW HPLC column equilibrated with 50 mM Tris-HCl pH 7.5, 150 mM NaCl. The material was eluted from the column at 0.5 ml/min, monitored at 220 nm and minute fractions collected.

In vitro phosphorylation assays. The auto-, trans-phosphorylation and in-blot renaturation studies were performed as in Example 1.

Antibodies and immunochemical techniques. Polyclonal antibodies were raised by in

chicken against a synthetic peptide (GPBPpep1) representing the sequence coded by the 78-bp exon (Genosys). Egg yolks were diluted 1:10 in water, the pH adjusted to 5.0. After 6 hours at 4 C, the solution was clarified by centrifugation (25 min at 10000 x g at 4°C) and the antibodies precipitated by adding 20 % (w/v) of sodium sulfate at 20.000 x g, 20°. The pellets were dissolved in PBS (1 ml per yolk) and used for immunohistochemical studies. The production of antibodies against GPBP/GPBP Δ 26 or against α 3(IV)NC1 domain are discussed above (see also 4, 13).

Sedimentation velocity. Determination of sedimentation velocities were performed in an Optima XL-A analytical ultracentrifuge (Beckman Instruments Inc.), equipped with a VIS-UV scanner, using a Ti60 rotor and double sector cells of Epon-charcoal of 12

mm optical path-length. Samples of ca. 400 µl were centrifuged at 30,000 rpm at 20°C and radial scans at 220 nm were taken every 5 min. The sedimentation coefficients were obtained from the rate of movement of the solute boundary using the program XLAVEL (supplied by Beckman).

Sedimentation equilibrium. Sedimentation equilibrium experiments were done as described above for velocity experiments with samples of 70 µl, and centrifuged at 8,000 rpm. The experimental concentration gradients at equilibrium were analyzed using the program EQASSOC (Beckman) to determine the corresponding weight average molecular mass. A partial specific volumes of 0.711 cm³/g for GPBP and 0.729 cm³/g for GPBPA26 were calculated from the corresponding amino acid compositions.

Physical methods and immunochemical techniques. SDS-PAGE and Western blotting were performed under reducing conditions as previously described (3). Immunohistochemistry studies were done on formalin fixed paraffin embedded tissues using the ABC peroxidase method (4) or on frozen human biopsies fixed with cold acetone using standard procedures for indirect immunofluorescence.

Two hybrid studies. Self-interaction studies were carried out in Saccharomyces cerevisiae (HF7c) using pGBT9 and pGAD424 (CLONTECH) to generate GAL4 binding and activation domain-fusion proteins, respectively. Interaction was assessed following the manufacture's recommendations. β -galactosidase activity was assayed with X-GAL (0.75 mg/ml) for in situ and with ortho-nitrophenyl β -D galactopyranoside (0.64 mg/ml) for the in-solution determinations.

RESULTS

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Identification of two spliced GPBP variants. To characterize the GPBP species in normal human tissues, we coupled reverse transcription to a polymerase chain reaction (RT-PCR) on total RNA from different tissues, using specific oligonucleotides that flank the full open reading frame of GPBP. A single cDNA fragment displaying lower size than expected was obtained from skeletal muscle-derived RNA (Fig.8A), and from kidney, lung, skin, or adrenal gland-derived RNA (not shown). By combining nested PCR re-amplifications and endonuclease restriction mapping, we determined that all the RT-PCR products corresponded to the same molecular species (not shown). We fully sequenced the 2.2-Kb of cDNA from human

muscle and found it identical to HeLa-derived material except for the absence of 78nucleotides (positions 1519-1596), which encode a 26-residues motif (amino acids 371-396) (Fig. 8B). We therefore named this more common isoform of GPBP as GPBPΔ26.

To investigate whether the 78-bp represent an exon skipped transcript during pre-mRNA processing, we used this cDNA fragment to probe a human-derived genomic library and we isolated a ~14-Kb clone. By combining Southern blot hybridization and PCR, the genomic clone was characterized and a contiguous DNA fragment of 12482-bp was fully sequenced (SEQ ID 25). The sequence contained (from 5' to 3'), 767-bp of intron sequence, a 93-bp exon, an 818-bp intron, the 78-bp exon sequence of interest, a 9650-bp intron, a 96-bp exon and a 980-bp intron sequence (Fig. 8C). The exon-intron boundaries determined by comparing the corresponding DNA and cDNA sequences meet the canonical consensus for 5' and 3' splice sites (Fig 8C) (5), thus confirming the exon nature of the 78-bp sequence. The GPBP gene was localized to chromosome 5q13 by fluorescence in situ hybridization (FISH) using the genomic clone as a probe (not shown).

The relative expression of GPBP in human-derived specimens was assessed by Northern blot analysis, using either the 78-bp exon or a 260-bp cDNA representing the flanking sequence of 78-bp (103-bp 5' and 157-bp 3') present in both GPBP and GPBP Δ 26 (Fig. 9). The 78-bp containing the molecular species of interest were preferably expressed in striated muscle (both skeletal and heart) and brain, and poorly expressed in placenta, lung and liver. In contrast to GPBP Δ 26, the GPBP was expressed at very low levels in kidney, pancreas and cancer cell lines.

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All the above indicates that GPBP is expressed at low levels in normal human tissues, and that the initial lack of detection by RT-PCR of GPBP can be attributed to a preferential amplification of the more abundant GPBPΔ26. Indeed, the cDNA of GPBP could be amplified from human tissues (skeletal muscle, lung, kidney, skin and adrenal gland) when the specific RT-PCR amplifications were done using 78-bp exon-specific oligonucleotides (not shown). This also suggests that GPBPΔ26 mRNA is the major transcript detected in Northern blot studies when using the cDNA probe representing both GPBP and GPBPΔ26.

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Recombinant expression and functional characterization of GPBPA26. To investigate whether the absence of the 26-residue serine-rich motif would affect the biochemical properties of GPBP, we expressed and purified both isoforms (rGPBP and rGPBPA26), and assessed their auto- and trans-phosphorylation activities (Fig. 10). As reported above for rGPBP (see also 4), rGPBPA26 is purified as a single major polypeptide and several related minor products (Fig.10 A). However, the number and relative amounts of the derived products vary compared to rGPBP, and they display M, on SDS-PAGE that cannot be attributed simply to the 26-residue deletion. This suggests that the 26-residue motif has important structural and functional consequences that could account for the reduced in-solution auto- and trans-phosphorylation activities displayed by rGPBPA26 (Fig. 10B). Interestingly, the differences in specific activity shown in the in-solution assays were not evident when autophosphorylation was assessed in-blot after SDS-PAGE and renaturation, suggesting that the 26-residue motif likely has important functional consequences at the quaternary structure level. Renaturation studies further showed that phosphate transfer activities reside in the major polypeptides representing the proposed open reading frames, and are not detectable in derived minor products.

rGPBP and rGPBP-26 exist as very active high molecular weight aggregates. Gel filtration analysis of affinity-purified rGPBP or rGPBPΔ26 yielded two chromatographic peaks (I and II), both displaying higher MW than expected for the individual molecular species, as determined by SDS-PAGE studies (89 kDa and 84 kDa, respectively) (Fig. 11). The bulk of the recombinant material eluted as a single peak between the 158 kDa and the 669 kDa molecular weight markers (peak II), while limited amounts of rGPBP and only traces of rGPBPΔ26 eluted in peak I (>1000 kDa). Aliquots of fractions representing each chromatographic profile were subjected to SDS-PAGE and stained, or incubated in the presence of ³²P[γ] ATP, and analyzed by immunoblot and autoradiography. Along with the major primary polypeptide, every chromatographic peak contained multiple derived products of higher or lower sizes indicating that the primary polypeptide associates to form high molecular weight aggregates that are stabilized by covalent and non-covalent bonds (not shown). The kinase activity also exhibited two peaks coinciding with the chromatographic profiles.

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However, peak I showed a much higher specific activity than peak II, indicating that these high molecular weight aggregates contained a much more active form of the kinase. Equal volumes of rGPBP fractions number 13 and 20 exhibited comparable phosphorylating activity, even though the protein content is approximately 20 times lower in fraction 13, as estimated by Western blot and Coomasie blue staining (Fig. 11A). The specific activities of rGPBP and rGPBPA26 at peak II are also different, and are consistent with the studies shown for the whole material, thus supporting the hypothesis that the presence of the 26-rediue serine-rich motif renders a more active kinase. These results also suggest that both rGPBP and rGPBPΔ26 exist as oligomers under native conditions, and that both high molecular weight aggregate formation and specific activity are greatly dependent on the presence of the 26-residue serine-rich motif. Analytical centrifugation analysis of rGPBP revealed that peak I contained large aggregates (over 10⁷ Da). Peak II of rGPBP contained a homogenous population of 220 ± 10 kDa aggregates, likely representing trimers with a sedimentation coefficient of 11S. Peak Π of rGPBPΔ26 however consisted of a more heterogenous population that likely contains several oligomeric species. The main population (ca. 80%) displayed a weight average molecular mass of 310 ± 10 kDa and a coefficient of sedimentation of 14S.

GPBP and GPBP Δ 26 self-interact in a yeast two-hybrid system. To assess the physiological relevance of the self-aggregation, and to determine the role of the 26-residue motif, we performed comparative studies using a two-hybrid interaction system in yeast. In this type of study, the polypeptides whose interaction is under study are expressed as a part of a fusion protein containing either the activation or the binding domains of the transcriptional factor GAL4. An effective interaction between the two fusion proteins through the polypeptide under study would result in the reconstitution of the transcriptional activator and the subsequent expression of the two reporter genes, Lac Z and His3, allowing colony color detection and growth in a His-defective medium, respectively. We estimated the intensity of interactions by the growth-rate in histidine-defective medium, in the presence of different concentrations of a competitive inhibitor of the His3 gene product (3-AT), and a quantitative colorimetric liquid β -galactosidase assay. A representative experiment is presented in Fig. 12. When

assaying GPBPΔ26 for self-interaction, a significant induction of the reporter genes was observed, while no expression was detectable when each fusion protein was expressed alone or with control fusion proteins. The insertion of the 26-residue motif in the polypeptide to obtain GPBP resulted in a notable increase in polypeptide interaction. All of the above data indicate that GPBPΔ26 self-associates *in vivo*, and that the insertion of the 26-residues into the polypeptide chain yields a more interactive molecular species.

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GPBP is highly expressed in human but not in bovine and murine glomerulus and alveolus. We have shown that GPBP/GPBP 26 is preferentially expressed in human cells and tissues that are commonly targeted in naturally occurring autoimmune responses. To specifically investigate the expression of GPBP, we raised polyclonal antibodies against a synthetic peptide representing the 26-residue motif characteristic of this kinase isoform, and used it for immunohistochemical studies on frozen or formalin fixed paraffin embedded human tissues (Fig 13). In general, these antibodies showed more specificity than the antibodies recognizing both isoforms for the tissue structures that are target of autoimmune responses such as the biliary ducts, the Langerhans islets or the white matter of the central nervous system (not shown). Nevertheless, the most remarkable finding was the presence of linear deposits of GPBP-selective antibodies around the small vessels in every tissue studied (A), suggesting that GPBP is associated with endothelial basement membranes. Consequently, at the glomerulus, the anti-GPBP antibodies displayed a vascular pattern closely resembling the glomerular basement membrane staining yielded either by monoclonal antibodies specifically recognizing the a3(IV)NC1 (compare 13B with 13C and 13D), or by circulating GP autoantibodies (compare 13E and 13F). These observations further supported the initial observation that GPBP is expressed in tissue structures targeted in natural autoimmune responses, suggesting that the expression of GPBP is a risk factor and makes the host tissue vulnerable to an autoimmune attack.

To further assess this hypothesis, we investigated the presence of GPBP and GPBPΔ26 in the glomerulus of two mammals that naturally do not undergo GP disease compared to human (Fig.14). GPBP-specific antibodies failed to stain the glomerulus of both bovine or murine specimens (compare 14A with 14B and 14C) while antibodies

recognizing the N-terminal sequence common to both GPBP and GPBPΔ26 stained these structures in all three species, although with different distributions and intensities (14D-14F). In bovine renal cortex, GPBPΔ26 was expressed at a lower rate than in human, but showed similar tissue distribution. In murine samples, however, GPBPΔ26 displayed a tissue distribution closely resembling that of GPBP in human glomerulus. Similar results were obtained when studying the alveolus in the three different species (not shown). To rule out that the differences in antibody detection was due to primary structure differences rather than to a differential expression, we determined the corresponding primary structures in these two species by cDNA sequencing. Bovine and mouse GPBP (SEQ ID NOS:3-6 and 9-12) displayed an overall identity with human material of 97.9% and 96.6% respectively. Furthermore, the mouse 26-residue motif was identical to human while bovine diverged only in one residue. Finally, and similarly to human, we successfully amplified GPBP cDNA from mouse or bovine kidney total RNA using oligonucleotides specific for the corresponding 78-bp exons, indicating that GPBP is expressed at very low levels not detectable by immunochemical techniques.

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GPBP is highly expressed in several autoimmune conditions. We analyzed several tissues from different GP patients by specific RT-PCR to assess GPBP/GPBPΔ26 mRNA levels. As in control kidneys, the major expressed isoform in GP kidneys was GPBPΔ26. However, in the muscle of one of the patients, GPBP was preferentially expressed, whereas GPBPΔ26 was the only isoform detected in control muscle samples (Fig. 15 A). Since we did not have kidney samples from this particular patient, we could not assess GPBP/GPBPΔ26 expression in the corresponding target organ. For similar reasons, we could not assess GPBP/GPBPΔ26 levels in the muscle of the patients in which kidneys were studied. Muscle cells express high levels of GPBP/GPBPΔ26 (see Northern blot in Fig. 9), and they comprise the bulk of the tissue. In contrast, the expression of GPBP/GPBPΔ26 in the kidney was much less, and the glomerulus was virtually the only kidney structure expressing the GPBP isoform (see Fig. 13). The glomerulus is a relatively less abundant structure in kidney than the myocyte is in muscle, and the glomerulus is the structure targeted by immune attack in GP pathogenesis. These factors, together with the preferential amplification of the more

abundant and shorter messages when performing RT-PCR studies, could account for the lack of detection of GPBP in both normal and GP kidneys, thus precluding the assessment of GPBP expression at the glomerulus during pathogenesis. Nevertheless, the increased levels of GPBP in a GP patient suggest that GPBP/GPBPΔ26 expression is altered during GP pathogenesis, and that augmented GPBP expression has a pathogenic significance in GP disease.

To investigate the expression of GPBP and GPBPΔ26 in autoimnune pathogenesis, we studied cutaneous autoimmune processes and compared them with control samples representing normal skin or non-autoimmune dermatitis (Fig. 15). Control samples displayed a limited expression of GPBP in the most peripheral keratinocytes (15B, 15E), while keratinocytes expanding from stratum basale to corneum expressed abundant GPBP in skin affected by systemic lupus erythematosus (SLE) (15C, 15F) or lichen planus (15D, 15G). GPBP was preferentially expressed in cell surface structures that closely resembled the blebs previously described in cultured keratinocytes upon UV irradiation and apoptosis induction (6). In contrast, antibodies recognizing both GPBP and GPBPΔ26 yielded a diffuse cytosolic pattern through the whole epidermis in both autoimmune affected or control samples (not shown). These data indicate that in both control and autoimmune-affected keratinocytes, GPBPA26 was expressed at the cytosol and that the expression did not significantly vary during cell differentiation. In contrast, mature keratinocytes were virtually the only GPBP expressing cells. However, bleb formation and expression of GPBP was observed in the early stages of differentiation in epidermis affected by autoimmune responses (15C, 15D, 15F, 15G). This further supports previous observations indicating that aberrant apoptosis at the basal keratinocytes is involved in the pathogenesis of autoimmune processes affecting skin (7), and suggests that apoptosis and GPBP expression are linked in this human cell system.

DISCUSSION

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Alternative pre-mRNA splicing is a fundamental mechanism for differential gene expression that has been reported to regulate the tissue distribution, intracellular localization, and function of different protein kinases (8-11). In this regard, and closely

resembling GPBP, B-Raf exists as multiple spliced variants, in which the presence of specific exons renders more interactive, efficient and oncogenic kinases (12).

Although it is evident that rGPBP \(\Delta 26 \) still bears the uncharacterized catalytic domain of this novel kinase, both auto- and trans-phosphorylating activities are greatly reduced when compared to rGPBP. Gel filtration and two hybrid experiments provide some insights into the mechanisms that underlie such a reduced phosphate transfer activity. About 1-2% of rGPBP is organized in very high molecular weight aggregates that display about one third of the phosphorylating activity of rGPBP, indicating that high molecular aggregation renders more efficient quaternary structures. Recombinant GPBPA26, with virtually no peak I material, consistently displayed a reduced kinase activity. However, aggregation does not seem to be the only mechanism by which the 26-residues increases specific activity, since the rGPBPΔ26 material present in peak II also shows a reduced phosphorylating activity when compared to homologous fractions of rGPBP. One possibility is that rGPBP-derived aggregates display higher specific activities because of quaternary structure strengthening caused by the insertion of the 26-residue motif. The oligomers are kept together mainly by very strong non-covalent bonds, since the bulk of the material appears as a single polypeptide in non-reducing SDS-PAGE, and the presence of either 8 M urea or 6 M guanidine had little effect on chromatographic gel filtration profiles (not shown). How the 26-residue motif renders a more strengthened and active structure remains to be clarified. Conformational changes induced by the presence of an exon encoded motif that alter the activation status of the kinase have been proposed for the linker domain of the Src protein (24) and exons 8b and 10 of B-Raf (12). Alternatively, the 26-residue motif may provide the structural requirements such as residues whose phosphorylation may be necessary for full activation of GPBP.

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We have reported (13) that the primary structure of the GP antigen ($\alpha 3$ (IV)NC1) is the target of a complex folding process yielding multiple conformers. Isolated conformers are non-minimum energy structures specifically activated by phosphorylation for supramolecular aggregation and likely quaternary structure formation. In GP patients, the $\alpha 3$ (IV)NC1 shows conformational alterations and a reduced ability to mediate the disulfide stabilization of the collagen IV network. The GP antibodies, in turn, demonstrate

stronger affinity towards the patient $\alpha 3(IV)NC1$ conformers, indicating that conformationally altered material caused the autoimmune response. Therefore, it seems that in GP disease an early alteration in the conforming process of the $\alpha 3(IV)NC1$ could generate altered conformers for which the immune system is not tolerant, thus mediating the autoimmune response.

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Other evidence (Raya et al., unpublished results) indicates that phosphorylation is the signal that drives the folding of the $\alpha 3(IV)NC1$ into non-minimum energy ends. In this scenario, three features of the human α3(IV)NC1 system are of special pathogenic relevance when compared to the corresponding antigen systems from species that, like bovine or murine, do not undergo spontaneous GP disease. First, the N-terminus of the human \alpha3(IV)NC1 contains a motif that is phosphorylatable by PKA and also by GPBP (see above, and also 2-4). Second, the human gene generates multiples alternative products by alternative exon splicing (14,15). Exon skipping generates alternative products with divergent C-terminal ends that up-regulate the in vitro PKA phosphorylation of the primary \(\alpha(\text{IV})\)NC1 product (See below Example 3). Third, the human GPBP is expressed associated with glomerular and alveolar basement membranes, the two main targets in GP disease. The phosphorylation-dependent conforming process is also a feature of non-pathogenic NC1 domains (13), suggesting that the phosphorylatable N-terminus, the alternative splicing diversification, and the expression of GPBP at the glomerular and alveolar basement membranes, are all exclusively human features that place the conformation process of $\alpha 3(IV)NC1$ in a vulnerable condition. The four independent GP kidneys studied expressed higher levels of GP antigen alternative products (15; Bernal and Saus, unpublished results), and an augmented expression of GPBP were found in a GP patient (see above). Both increased levels of alternative GP antigen products and GPBP are expected to have consequences in the phosphorylationdependent conformational process of the $\alpha 3(IV)NC1$, and therefore with pathogenic potential.

GPBP is highly expressed in skin targeted by natural autoimmune responses. In the epidermis, GPBP is associated with cell surface blebs characteristic of the apoptosis-mediated differentiation process that keratinocytes undergo during maturation from basale to corneum strata (22, 23). Keratinocytes from SLE patients

show a remarkably heightened sensitivity to UV-induced apoptosis (6, 18, 20), and augmented and premature apoptosis of keratinocytes has been reported to exist in SLE and dermatomyositis (7). Consistently, we found apoptotic bodies expanding from basal to peripheral strata of the epidermis in several skin autoimmune conditions including discoid lupus (not shown), SLE and lichen planus. Autoantigens, and modified versions thereof are clustered in the cell surface blebs of apoptotic keratinocytes (6,18,20). Apoptotic surface blebs present autoantigens (21), and likely release modified versions to the circulation (16-20). It has been suggested that the release of modified autoantigens from apoptotic bodies could be the immunizing event that mediates systemic autoimmune responses mediating SLE and scleroderma (18,19).

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Our evidence indicates that both GPBP and GPBPA26 are able to act in vitro as protein kinases, with GPBP being a more active isoform than GPBPA26. Furthermore, recombinant material representing GPBP or GPBPΔ26 purified from yeast or from human 293 cells contained an associated proteolytic activity that specifically degrades the \(\alpha \) (IV)NC1 domain (unpublished results). The proteolytic activity operates on α3(IV)NC1 produced in an eukaryotic expression system, but not on recombinant material produced in bacteria (unpublished results), indicating that \(\alpha3(IV)NC1\) processing has some conformational or post-translational requirements not present in prokaryotic recombinant material. Finally, it has been reported that several autoantigens undergo phosphorylation and degradation in apoptotic keratinocytes (20). While not being limited to an exact mechanism, we propose, in light of all of the above data, that the machinery assembling GPBP at the apoptotic blebs likely performs a complex modification of the autoantigens that includes phosphorylation, conformational changes and degradation. Accordingly, recombinant protein representing autoantigens in SLE (P1 ribosomal phosphoprotein and Sm-D1 small nuclear ribonucleoproteins) and in dermatomyositis (hystidil-tRNA synthetase) were in vitro substrates of GPBP (unpublished results).

The down-regulation in cancer cell lines of GPBP, suggest that the cell machinery harboring GPBP/GPBPA26 is likely involved in signaling pathways inducing programmed cell death. The corresponding apoptotic pathway could be up regulated during autoimmune pathogenesis to cause an altered antigen presentation in

individuals carrying specific MHC haplotypes; and down regulated during cell transformation to prevent autoimmune attack to the transformed cells during tumor growth.

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Example 3. Regulation of Human Autoantigen Phosphorylation by Exon Splicing

20 INTRODUCTION

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In GP disease, the immune system attack is mediated by autoantibodies against the non-collagenous C-terminal domain (NC1) of the α 3 chain of collagen IV (the GP antigen) (1). The N-terminus of the human α 3(IV)NC1 contains a highly divergent and hydrophilic region with a unique structural motif, KRGDS⁹, that harbors a cell adhesion signal as an integral part of a functional phosphorylation site for type A protein kinases (2,3). Furthermore, the gene region encoding the human GP antigen characteristically generates multiple mRNAs by alternative exon splicing (4,5). The alternative products diverge in the C-terminal ends and all but one share the N-terminal KRGDS⁹ (4,5).

Multiple sclerosis (MS) is an exclusive human neurological disease characterized by the presence of inflamatory demyelization plaques at the central nervous system. (6). Several evidences indicate that this disease is caused by an autoimmune attack mediated by cytotoxic T cells towards specific components of the white matter including the myelin basic protein (MBP) (7, 8). In humans, the MBP gene generates four products (MBP, MBPΔII, MBPΔV and MBPΔII/V) that result from alternative exon splicing during premRNA processing (9). Among these, MBPΔII is the more abundant form in the mature central nervous system, while MBP form containing all the exons is virtually absent (9).

Several biological similarities exist between the autoimme responses mediating GP disease and MS, namely: 1) both are human exclusive diseases and typically initiate after a viral flu-like disease; 2) a strong linkage exists to the same haplotype of the HLA-DR region of the class II MHC; 3) several products are generated by alternative splicing; and 4) the death of a MS patient by GP disease has recently been reported (10).

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MATERIALS AND METHODS

Synthetic polymers: GPAIII derived peptide, QRAHGQDLDALFVKVLRSP (SEQ ID NO:43) and GPAIII/IV/V derived peptide, QRAHGQDLESLFHQL (SEQ ID NO:44) were synthesized using either Boc- (MedProbc) or Fmoc- (Chiron, Lipotec) chemistry.

Plasmid construction and recombinant expression.

GP derived material: The constructs representing the different GP-spliced forms were obtained by subcloning the cDNAs used elsewhere to express the corresponding recombinant proteins (5) into the BamHI site of a modified pET15b vector, in which the extraneous vector-derived amino-terminal sequence except for the initiation Met was eliminated. The extra sequence was removed by cutting the vector with NcoI and Bam HI, filling-in of the free ends with Klenow, and re-ligation. This resulted in the reformation of both restriction sites and placed the BamHI site immediately downstream of the codon for the amino-terminal Met.

The recombinant proteins representing GP or GPAV (SEQ ID NO:46) were purified by precipitation (5). Bacterial pellets containing the recombinant proteins representing GPAIII (SEQ ID NO:48) or GPAIII/IV/V (SEQ ID NO:50) were dissolved by 8 M urea in 40 mM Tris-HCl pH 6.8 and sonication. After centrifugation at 40,000 x g the supernatants were passed through a 0.22 µm filter and applied to resource Q column for FPLC. The effluent was acidified to pH 6 with HCl and applied to a resource S column previously equilibrated with 40 mM MES pH 6 for a second FPLC

purification. The material in the resulting effluent was used for in vitro phosphorylation.

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MBP-derived material: cDNA representing human MBPΔII (SEQ ID NO:51) was obtained by RT-PCR using total RNA from central nervous system. The cDNA representing human MBP was a generous gift from C. Campagnoni (UCLA). Both fragments were cloned into a modified version of pHIL-D2 (Invitrogen) containing a 6xHis-coding sequence at the C-terminus to generate pHIL-MBPΔII-His and pHIL-MBP-His, respectively. These plasmids were used for recombinant expression in *Pichia pastoris* as described in (12). Recombinant proteins were purified using immobilized metal affinity chromatography (TALON resin, CLONTECH) under denaturant conditions (8M urea) and eluted with 300 mM imidazole following manufacturers' instructions. The affinity-purified material was then renatured by dilution into 80 volumes of 50 mM Tris-HCl pH 8.0, 10 mM CHAPS, 400 mM NaCl, 2 mM DTT, and concentrated 50 times by ultrafiltration through a YM10-type membrane (AMICON). The Ser to Ala mutants were produced by site-directed mutagenesis over native sequence-containing constructs using transformer mutagenesis kit from CLONTECH and the resulting proteins were similarly produced.

Phosphorylation studies. Phosphorylation studies were essentially done as described above (see also 3 and 12). In some experiments, the substrates were in-blot renatured and then, phosphorylated for 30 min at room temperature by overlaying 100 μ l of phosphorylation buffer containing 0.5 μ g of rGPBP. Digestion with V8 endopeptidase and immunoprecipitation were performed as described in (3).

Antibody production. Synthetic peptides representing the C-terminal divergent ends of GPΔIII or GPΔIII/IV/V comprised in SEQ ID NO:43 or SEQ ID NO:44 respectively were conjugated to a cytochrome C, BSA or ovoalbumine using a glutaraldehyde coupling standard procedure. The resulting protein conjugates were used for mouse immmunization to obtain polyclonal antibodies specific for GPΔIII and monoclonal antibodies specific for GPΔIII/IV/V (Mab153). To obtain monoclonal antibodies specific for GPΔV (Mab5A) mouse were immunized using recombinant bacterial protein representing the corresponding alternative form comprising the SEQ ID NO:50. The production of monoclonal (M3/1, P1/2) or polyclonal (anti-GPpcp1)

antibodies against SEQ ID NO: 26 which represents the N-terminal region of the GP alternative forms have been previously described (3,5).

Boc-based peptide synthesis.

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Assembling. The peptide was assembled by stepwise solid phase synthesis using a Boc-Benzyl strategy. The starting resin used was Boc-Pro-PAM resin (0.56 meq/g, batch R4108). The deprotection /coupling procedure used was: TFA (1x1min) TFA (1x3 min) DCM (flow flash) Isopropylalcohol (1x30 sec) DMF (3 x 1 min) COUPLING/DMF (1 x10 min) DMF (1x1 min) COUPLING/DMF (1x 10 min) DMF (2x 1min) DCM (1x 1min). For each step 10 ml per gram of peptide-resin were used. The coupling of all amino acids (fivefold excess) was performed in DMF in the presence of BOP, Hobt and DIEA. For the synthesis the following side-chain protecting groups were used: benzyl for serine; 2 chlorobenzyloxycarbonyl for lysine; cyclohexyl for aspartic and glutamic acid; tosyl for histidine and arginine.

Cleavage. The peptide was cleaved from the resin and fully deprotected by a treatment with liquid Hydrogen Fluoride (HF): Ten milliliters of HF per gram of peptide resin were added and the mixture kept at 0° C for 45 min in the presence of percesol as scavengers. After evaporation of the HF, the crude reaction mixture is washed with ether, dissolved in TFA, precipitated with ether and dried.

Purification. Stationary phase: Silica C18, 15 μm, 120 A; Mobile phase: solvent A: water 0.1% TFA and solvent B: acetonitrile /A, 60/40 (v/v); Gradient: linear from 20 to 60% B in 30 min; Flow rate: 40 ml/min; and detection was U.V (210 nm). Fractions with a purity higher than 80% were pooled and lyophilized. Control of purity and identity was performed by analytical HPLC and ES/MS. The final product had 88% purity and an experimental molecular weight of 2192.9.

Fmoc-based peptide synthesis.

Assembling. The peptides were synthesized by stepwise linear solid phase on Pro-clorotrityl-resin (0.685 meq/g) with standard Fmoc/tBu chemistry. The deprotection /coupling procedure used was: Fmoc aa (0.66 g) HOBt (0.26 g) DIPCDI (0.28 ml) for 40 min following a control by Kaiser test. If the test was positive the time was extended until change to negative. Then DMF (31 min), piperidine/DMF 20% (11 min) piperidine/DMF 20% (15 min) and DMF (41 min). Side chain protectors were:

Pmc (pentamethylcromane sulfonyl) for arginine, Bcc (tert-butoxycarbonyl) for lysine, tBu (tert-butyl) for aspartic acid and for serine and Trl (trityl) for histidine.

Cleavage. The peptide was cleaved and fully deprotected by treatment cleavage with TFA/water 90/10. Ten milliliters of TFA solution per gram of resin were added. Water acts as scavenger. After two hours, resin was filtered and the resulting solution was precipitated five times with cold diethylether. The final precipitated was dried.

Purification. Stationary phase: Kromasil C18 10 μm; Mobile phase: solvent A: water 0.1% TFA and solvent B: acetonitrile 0.1% TFA; Isocratic: 28% B; Flow rate: 55 ml/min; Detection: 220 nm. Fractions with the higher purity were pooled and lyophilized, and a second HPLC purification round performed. Control of purity and identity was performed by analytical HPLC and ES/MS. The final product had 97% purity and an experimental molecular weight of 2190.9.

RESULTS

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Regulation of the phosphorylation of the human GP antigen by alternative splicing. We produced bacterial recombinant proteins representing the primary antigen (GP) or the individual alternative products GPAV (SEQ ID NO:46), GPAIII (SEQ ID NO:48) and GPAIII/IV/V (SEQ ID NO:50), and we tested their ability to be phosphorylated by PKA (Figure 16, left panel). Using standard ATP concentrations (150 uM), all four recombinant antigens were phosphorylated but to very different extents. The alternative forms incorporated 32P more efficiently than the primary GP antigen, suggesting that they are better substrates. Because these antigens are expected to be in the extracellular compartment, we also assayed their phosphorylatability with more physiological ATP concentrations (0.1-0.5 µM). Under these conditions, the differences in ³²P incorporation between the primary and alternative products were more evident, indicating that at low ATP concentrations the primary GP antigen was a very poor substrate for the kinase. Among the three PKA phosphorylation sites present in the GP antigen, the N-terminal Ser9 and Ser26 are the major ones, and are common to all the Accordingly, the differences observed in alternative products assayed (3,5). phosphorylation for the full polypeptides also existed among the individual N-terminal regions, as determined after specific V8 digestion and immunoprecipitation (not shown). This strongly suggests that differences in phosphorylation might be due to the presence of different C-terminal sequences in the alternative products. Since GPAIII and GPAIII/IV/V displayed significantly higher ³²P incorporation rates than GPAV, and they have shorter divergent C-terminal regions (5), we used synthetic peptides individually representing these C-terminal sequences (SEQ ID NO: 43, SEQ ID NO:44) to further examine their regulatory roles in the in vitro phosphorylation of the native antigen. Collagen IV is a trimeric molecule comprised of three interwoven a chains. In basement membranes, two collagen IV molecules assemble through their NC1 domains to yield a hexameric NC1 structure that can be solubilized by bacterial collagenase digestion (1). Dissociation of the hexamer structure releases the GP antigen in monomeric and disulfide-related dimeric forms (1). For the following set of experiments, we carried out phosphorylations in the presence of low, extracellular-like ATP concentrations using both monomeric or hexameric native GP antigen (Figure 16, right panel). The presence of each specific peptide but not control peptides (not shown) induced the phosphorylation of a single polypeptide displaying an apparent MW of 22 kDa. By specific V8 digestion and immunoprecipitation, the corresponding polypeptide has been identified as the 22 kDa conformer of the $\alpha 3(IV)NC1$, previously characterized and identified as the best substrate for the PKA (11).

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Regulation of the phosphorylation of the MBP by alternative splicing. The MBP contains at its N terminal region two PKA phosphorylation sites (Ser⁸, Ser⁵⁷) that are structurally similar to the N terminus site (Ser⁹) present in GP antigen products (Fig 17). The Ser⁸ site present in all the MBP proteins is located in a similar position than the Ser⁹ in the GP-derived polypeptides. In addition, in the MBP and GPΔIII Ser⁸ and Ser⁹ respectively are at a similar distance in the primary structures of a highly homologous motif present in the corresponding exon II (bend arrow in Fig 17). The GPΔIII-derived motif coincides with the C terminal divergent region that up-regulates PKA phosphorylation of Ser⁹ in the GP antigen system (Fig. 16). The regulatory-like sequence in MBP is located at exon II and its presence in the final products depends on an alternative exon splicing mechanism. Therefore, the MBP motif identified by structural comparison to GPΔIII may be also regulating PKA phosphorylation of Ser⁸. We produced recombinant proteins representing MBP and MBPΔII (SEQ ID NO:54) and the corresponding Ser to Ala mutants to knock-out each of the two PKA phosphorylation sites (Ser⁸ and Ser⁵⁷) present in exon I. Subsequently, we assessed its in vitro phosphorylation

by PKA (Fig. 18). MBPAII was a better substrate than MBP, and Ser⁸ was the major phosphorylation site, indicating that, similarly to GP antigenic system, alternative exon splicing regulates the PKA phosphorylation of specific sites located at the N-terminal region common to all the MBP-derived alternative forms.

In similar experiments assessing GPBP phosphorylation of the recombinant MBP proteins, GPBP preferentially phosphorylated MBP, while little phosphorylation of MBPAII was observed (Fig. 19). Furthermore, recombinant Ser to Ala mutants displayed no significant reduction in ³²P incorporation, indicating that GPBP phosphorylates MBP/MBPAII in an opposite way than PKA, and that these two kinases do not share major phosphorylation sites in MBP proteins.

From all these data we concluded that in the MBP system, alternative splicing regulates the phosphorylation of specific serines by either PKA or GPBP.

Synthetic peptides representing the C terminal region of GPΔIII influence GPBP phosphorylation. To assess the effect of the C terminal region of GPΔIII on GPBP activity, peptides representing this region were synthesized using two different chemistries (Boc or Fmoc), and separately added to a phosphorylation mixture containing GPBP (Fig. 20). Boc-based synthetic peptides positively influenced GPBP autophosphorylation while Fmoc-based inhibited GPBP autophosphorylation, suggesting that the regulatory sequences derived from the alternative products in either GP and MBP antigenic systems can influence the kinase activity of GPBP.

DISCUSSION

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We have shown that the $\alpha 3(IV)NC1$ domain undergoes a complex structural diversification by two different mechanism: 1) alternative splicing (4,5) and 2) conformational isomerization of the primary product (11). Both mechanisms generate products that are distinguished by PKA, indicating that PKA phosphorylation is a critical event in the biology of the $\alpha 3(IV)NC1$ domain. Phosphorylation guides at least in part the folding, but also the supramolecular assembly of the $\alpha 3(IV)NC1$ domain in the collagen IV network (11 and Raya et al. unpublished results). Altered conformers of the $\alpha 3(IV)NC1$ lead the autoimmune response mediating GP disease (11), suggesting that an alteration in antigen phosphorylation could be the primary event in the onset of

the disease. Accordingly, we have found increased expression levels of GPAIII in several GP kidneys (4 and Bernal and Saus, unpublished results), and an increased expression of GPBP has been detected in another Goodpasture patient (Fig. 15). Both increased expression of alternative GP antigen products and of GPBP are expected to have consequences in the phosphorylation steady state of $\alpha 3(IV)NC1$, and therefore in the corresponding conformational process. The discrimination among the different structural products by PKA strongly suggests that this kinase, or another structurally similar kinase, is involved in the physiological antigen conforming process, and that antigen phosphorylation by GPBP has a pathogenic significance. In pathogenesis, GPBP could be an intruding kinase, interfering in the phosphorylation-dependent conforming process. Accordingly, GPBP is expressed in tissue structures that are targeted by natural autoimmune responses, and an increased expression of GPBP is associated with several autoimmune conditions (See examples 1 and 2 above).

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An alternative splicing mechanism also regulates the PKA phosphorylation of specific serines in the MBP antigenic system. MBP is also a substrate for GPBP suggesting that GPBP may play a pathogenic role in multiple sclerosis, and other autoimmune responses.

All of the above data identify GPBP as a potential target for therapeutics in autoimmune disease. In Fig 20, we show that synthetic peptides representing the C terminal region of GPAIII (SEQ ID NO:43) modulate the action of GPBP in vitro, and therefore we identified this and related sequences as peptide-based compounds to modulate the activity of GPBP in vivo. The induction of GP antigen phosphorylation by PKA was achieved when using Boc-based peptides, but not when using similar Fmoc-based peptides. Furthermore, Boc- but not Fmoc-based peptides were in vitro substrates of PKA (not shown), indicating that important structural differences exist between both products. Since both products displayed no significant differences in mass spectrometry, one possibility is that the different deprotection procedure used may be responsible for conformational differences in the secondary structure that may be critical for biological activity. Accordingly, Boc-based peptide loses its ability to induce PKA upon long storage at low temperatures.

REFERENCES FOR EXAMPLE 3

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The present invention is not limited by the aforementioned particular preferred embodiments. It will occur to those ordinarily skilled in the art that various modifications may be made to the disclosed preferred embodiments without diverting from the concept of the invention. All such modifications are intended to be within the scope of the present invention.

I claim:

- 1. An isolated nucleic acid sequence comprising a sequence substantially similar to a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, and SEQ ID NO:25.
- 2. An isolated nucleic acid sequence comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, and SEQ ID NO:25.
- 3. An isolated nucleic acid comprising a sequence that encodes a polypeptide selected from the group consisting of GPBP, GPBPΔ26, and GPBPpep1, or fragments thereof.
 - 4. An isolated nucleic acid sequence comprising a sequence that encodes a protein sequence substantially similar to a protein sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, and SEQ ID NO:24
 - 5. An isolated nucleic acid sequence comprising a sequence that encodes a protein sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, and SEQ ID NO:24.
- 6. A recombinant expression vector comprising the isolated nucleic acid sequence of any one of claims 1-5.

- 7. A recombinant expression vector comprising an isolated nucleic acid sequence comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, and SEQ ID NO:25, or fragments thereof
- 8. A host cell transfected with the recombinant expression vector of claim 6 or 7.
- 9. A substantially purified polypeptide, comprising an amino acid sequence substantially similar to a sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, or peptide fragments thereof

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- 15 10. A substantially purified polypeptide, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, or peptide fragments thereof.
- 20 11. A substantially purified protein comprising a polypeptide selected from the group consisting of GPBP, GPBPΔ26, and GPBPpep1, or peptide fragments thereof.
 - 12. An antibody that selectively binds to the substantially purified protein or polypeptide of any one of claims 9-11.
 - 13. The antibody of claim 12, wherein the antibody is a polyclonal antibody.
 - 14. The antibody of claim 12, wherein the antibody is a monoclonal antibody.
- 30 15. A method for detecting the presence of a protein that is substantially similar to a protein selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID

NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, comprising

a) providing a protein sample to be screened;

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- b) contacting the protein sample to be screened with the antibody of any one of claims 12-14 under conditions that promote antibody-antigen complex formation; and
 - c) detecting the formation of antibody-antigen complexes, wherein the presence of the antibody-antigen complex indicates the presence of a protein that is substantially similar to a protein selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24.
- 16. The method of claim 15, wherein detecting comprises a method selected from the group consisting of immunolocalization, immunofluorescence analysis, Western blot analysis, ELISAs, and nucleic acid expression library screening.
 - 17. A method for detecting in a sample a sequence that is substantially similar to a nucleic acid selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, or SEQ ID NO:25, comprising contacting the sample with the isolated nucleic acid of any one of claims 1-5, or fragments thereof, and detecting complex formation, wherein complex formation indicates the presence in the sample of the sequence that is substantially similar to a nucleic acid selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEO ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, or SEQ ID NO:25.
 - 18. The method of claim 17, wherein the detecting is carried out by a method selected from the group consisting of hybridization, reverse transcription, PCR, coupled reverse transcription-PCR, Northern blotting, Southern blotting, and DNA library screening.

- A method for detecting an autoimmune condition in a patient, comprising
 -providing a tissue or body fluid sample from the patient;
- -providing a control tissue or body fluid sample in which no autoimmune condition is present; and
 - -detecting altered GPBP RNA or protein expression in the tissue or body fluid sample compared to the control sample, wherein an alteration in GPBP RNA or protein expression relative to the control indicates the presence of an autoimmune condition.
- 10 20. A method for detecting cells undergoing apoptosis or cancer transformation in a tissue or body fluid sample, comprising
 - -providing a tissue or body fluid sample from the patient;
 - -providing a normal control tissue or body fluid sample; and
- -detecting altered GPBP RNA or protein expression in the tissue or body fluid

 sample compared to the control sample, wherein an alteration in GPBP RNA or protein expression relative to the control indicates the presence of cells undergoing apoptosis or cancer transformation.
- 21. A method for treating a patient with an autoimmune disorder, comprising modifying the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1 in the patient with the autoimmune disorder.
- 22. A method for treating a patient with a tumor, comprising modifying the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1 in the patient with the tumor.
- 23. A method for preventing cell apoptosis, comprising modifying the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially
 30 similarly to GPBPpep1 in the cell.

24. The method of claim 21, 22, or 23 wherein alternative products of the Goodpasture antigen or of the myelin basic protein are used to modify the expression or activity of GPBP, GPBP Δ 26 or a protein comprising a polypeptide substantially similarly to GPBPpep1.

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- 25. The method of claim 21, 22, or 23 wherein nucleic acids comprising sequences substantially similar to SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO: 51, or SEQ ID NO:53 or fragments thereof are used to modify the expression or activity of GPBP, GPBP Δ 26 or a protein comprising a polypeptide substantially similarly to GPBPpep1.
- 26. The method of claim 21, 22, or 23 wherein polypeptides comprising sequences substantially similar to SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:54, or fragments thereof are used to modify the expression or activity of GPBP, GPBPΔ26 or a protein comprising a polypeptide substantially similarly to GPBPpep1.
- 27. An isolated nucleic acid sequence comprising a sequence that encodes a polypeptide substantially similar to an amino acid sequence selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, or peptide fragments thereof.
- 28. An isolated nucleic acid sequence comprising a sequence that encodes a polypeptide selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, and peptide fragments thereof.

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- 29. A recombinant expression vector comprising the isolated nucleic acid sequence of claim 27 or 28.
- 30. A host cell transfected with the recombinant expression vector of claim 29.

- 31. A substantially purified polypeptide, comprising an amino acid sequence substantially similar to a sequence selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, or peptide fragments thereof
- 5 32. A substantially purified polypeptide, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, or peptide fragments thereof.
- 33. An antibody that selectively binds to the substantially purified protein or polypeptide of claim 31 or 32.
 - 34. The antibody of claim 33, wherein the antibody is a polyclonal antibody.
 - 35. The antibody of claim 33, wherein the antibody is a monoclonal antibody.

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- 36. The method of claim 21, 22, or 23 comprising administering a substantially purified polypeptide substantially similar to a polypeptide selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:54, or fragments thereof, to modify the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1.
- 37. The method of claim 21, 22, or 23 comprising administering an isolated nucleic acid comprising sequences substantially similar to SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO: 51, or SEQ ID NO:53 or fragments thereof, or fragments thereof, to modify the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1.
- 38. A pharmaceutical composition, comprising an amount effective of a substantially purified polypeptide substantially similar to a polypeptide selected from the group consisting of SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:54, or fragments thereof, to modify the

expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1, and a pharmaceutically acceptable carrier.

39. A pharmaceutical composition, comprising an amount effective of a an isolated nucleic acid comprising sequences substantially similar to SEQ ID NO:45, SEQ ID NO:45, SEQ ID NO:47, SEQ ID NO:49, SEQ ID NO: 51, or SEQ ID NO:53 or fragments thereof, to modify the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1, and a pharmaceutically acceptable carrier.

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40. The method of claim 21, 22, or 23 comprising administering the pharmaceutical composition of claim 38 or 39 to modify the expression or activity of GPBP, GPBPΔ26, or a protein comprising a polypeptide substantially similarly to GPBPpep1.

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${\tt GCAGGAAGATGGCGGGGGGGGGGGGGGGGGGGGGGGGGG$	70
${\tt TCTTTTCCCTTTTCCCTATTTGAAATTGGCATCGAGGGGGCTAAGTTCGGGTGGCAGCGCCGGGCGGG$	140
$\tt CAACGCAGGGGTCACGGCGGCGGCGGCGGCGGCGGCTGACGGCTGGAAGGGTAGGCTTCATTCA$	210
CTCCTTCCTCGCTCCGCTCGGTGTCAGGCGCGGCGGCGGCGGGCG	280
${\tt TCCCCCCACACCGGAGCGGGCACTCTTCGCTTCGCCATCCCCCGACCCTTCACCCCGAGGACTGGGCGCCCCCCCC$	350
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	420 4
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	490 27
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	560 51
TCTGAGTTACTACAAATCTGAAGATGAAACAGAGTATGGCTGCAGAGGATCCATCTGTCTTAGCAAGGCT L S Y Y K S B D E T E Y G C R G S I C L S K A	630 74
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	700 97
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	770 121
TGGATCTGAATCCAGCTTGCGTCGACATGGCTCAATGGTGTCCCTGGTGTCTCGGAGCAAGTGGCTACTCT G S E S S L R R H G <u>S M V S L V S G A S G Y S</u>	840 144
GCAACATCCACCTCTTCATTCAAGAAAGGCCACAGTTTACGTGAGAAGTTGGCTGAAATGGAAACATTTA A T S T S S F K K G H S L R E K L A E M E T F	910 167
GAGACATCTTATGTAGACAAGTTGACACGCTACAGAAGTACTTTGATGCCTGTGCTGATGCTGTCTCTAA R D I L C R Q V D T L Q K Y F D A C A D A V S K	980 191
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	1050 214
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	1120 237
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	
TTGTATTGAACTAATGGTTAAACGTGAGGACAGCTGGCAGAAGAGACTGGATAAGGAAACTGAGAAGAAA C I E L M V K R E D <u>S W Q K R L D K E T E K K</u>	
AGAAGAACAGAGGAAGCATATAAAAATGCAATGACAGAACTTAAGAAAAAATCCCACTTTGGAGGACCAG R R T B B A Y K N A M T B L K K K S H F G G P	
ATTATGAAGAAGGCCCTAACAGTCTGATTAATGAAGAAGAGTTCTTTGATGCTGTTGAAGCTGCTCTTGA	

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FIG. 1

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CAGACAAGATAAAATAGAAGAACAGTCACAGAGTGAAAAAGGTGAGATTACATTGGCCTACATCCTTGCCCRQQDK1EEQQCTACATCCTTGCCC	1470 354
TCTGGAGATGCCTTTTCTTCTGTGGGGACACATAGATTTGTCCAAAAGCCCTATAGTCGCTCTTCCTCCASG D A F S S V G T H R F V Q K P Y S R S S S	1540 377
TGTCTTCCATTGATCTAGTCAGTGCCTCTGATGATGTTCACAGATTCAGCTCCCAGGTTGAAGAGATGGTM S S I D L V S A S D D V H R F S S Q V E E M $^{\circ}$	
GCAGAACCACATGACTTACTCATTACAGGATGTAGGCGGAGATGCCAATTGGCAGTTGGTTG	1680 424
GGAGAAATGAAGGTATACAGAAGAAGAAGTAGAAGAAAATGGGATTGTTCTGGATCCTTTAAAAGCTACCC G E M K V Y R R E V E E N G I V L D P L K A T	1750 447
$ \begin{array}{cccccccccccccccccccccccccccccccccccc$	3 1820 V 471
GGAAACAACTATAGAAAACTTTCATGTGGTGGAAACATTAGCTGATAATGCAATCATCATTATCAAACAETTTTTTCAAACAETTTTTTTCAAACAETTTTTTTT	494 494
CACAAGAGGGTGTGGCCTGCTTCTCAGCGAGACGTATTATATCTTTCTGTCATTCGAAAGATACCAGCCTH K R V W P A S Q R D V L Y L S V I R K I P A	r 1960 517
TGACTGAAAATGACCCTGAAACTTGGATAGTTTGTAATTTTTCTGTGGATCATGACAGTGCTCCTCTAA	
CAACCGATGTGTCCGTGCCAAAATAAATGTTGCTATGATTTGTCAAACCTTGGTAAGCCCACCAGAGGGANR R C V R A K I N V A M I C Q T L V S P P E G	A 2100 564
AACCAGGAAATTAGCAGGGACAACATTCTATGCAAGATTACATATGTAGCTAATGTGAACCCTGGAGGA N Q E I S R D N I L C K I T Y V A N V N P G G	r 2170 587
GGGCACCAGCCTCAGTGTTAAGGGCAGTGGCAAAGCGAGAGTATCCTAAATTTCTAAAACGTTTTACTTWAAPA SVLRAAVA KREYPKFLKRFT	
TTACGTCCAAGAAAAACTGCAGGAAAGCCTATTTTGTTCTAGTATTAACAGGTACTAGAAGATATGTT Y V Q E K T A G K P I L F	r 2310 624
TATCTTTTTTAACTTTATTTGACTAATATGACTGTCAATACTAAAATTTAGTTGTTGAAAGTATTTAC	r 2380
ATGTTTTT	2389

FIG. 1

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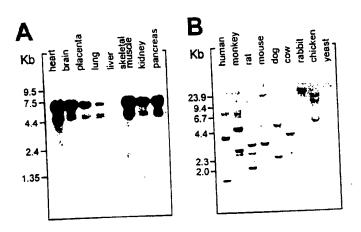


FIG. 2

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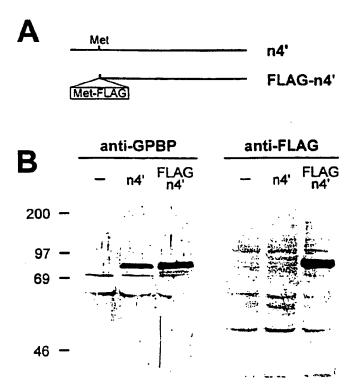
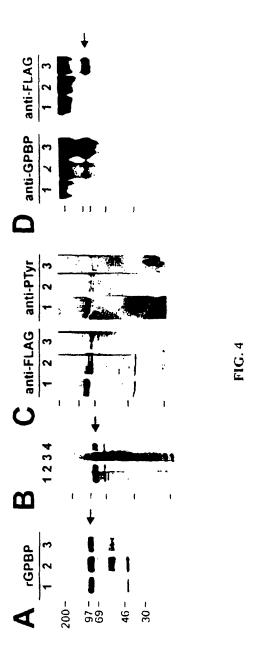
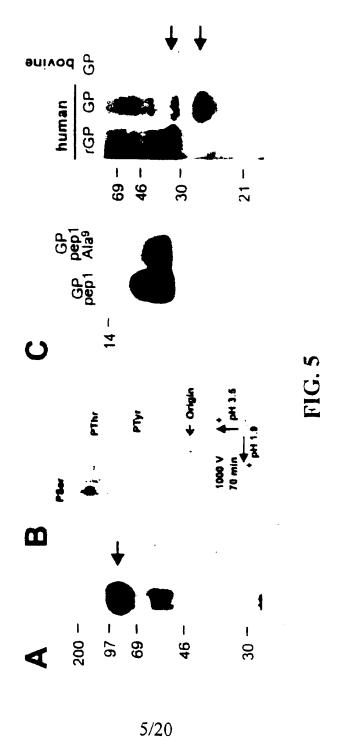


FIG. 3



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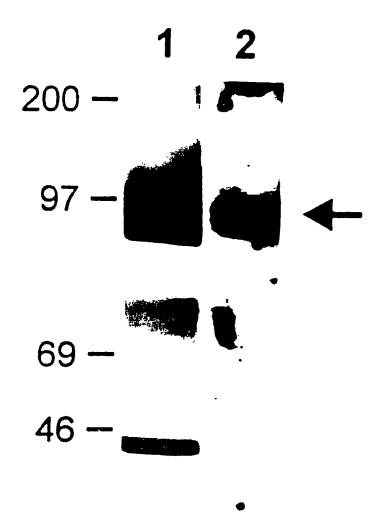


FIG. 6

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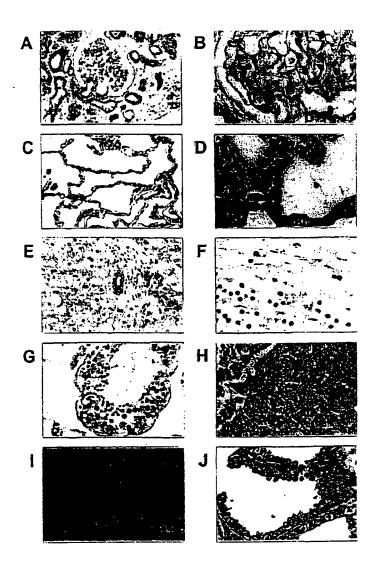
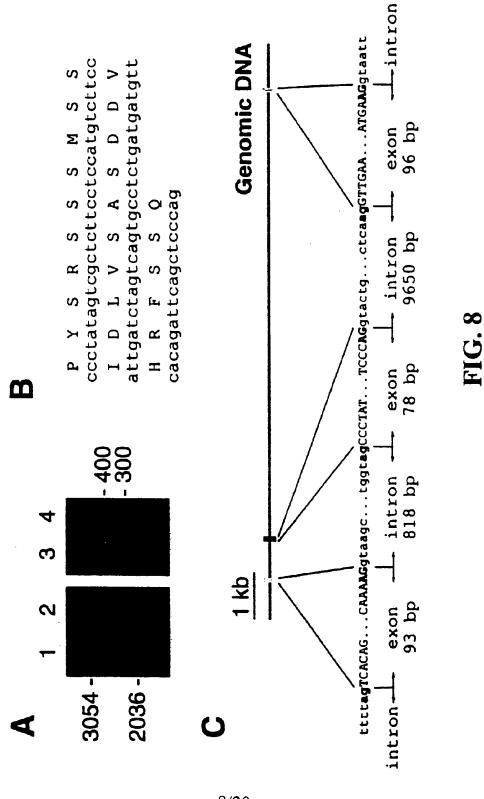


FIG. 7

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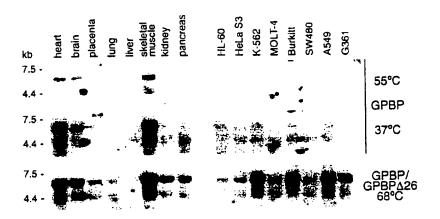
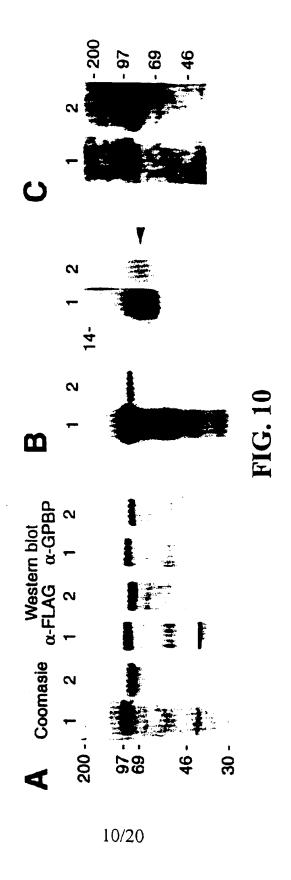


FIG. 9

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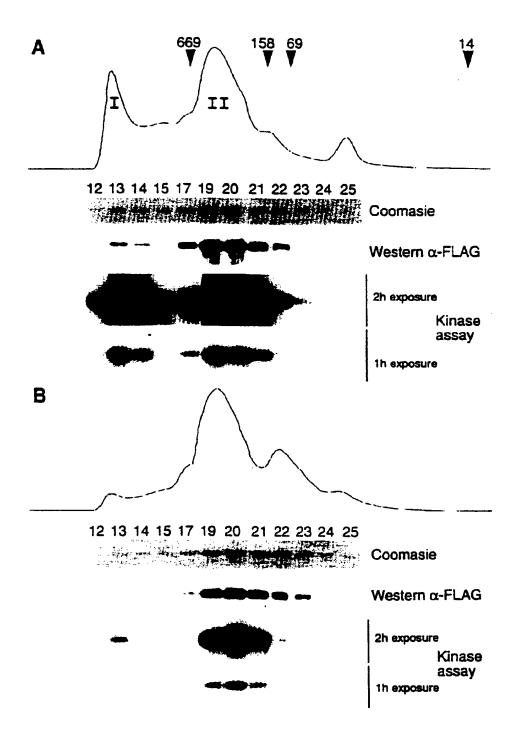
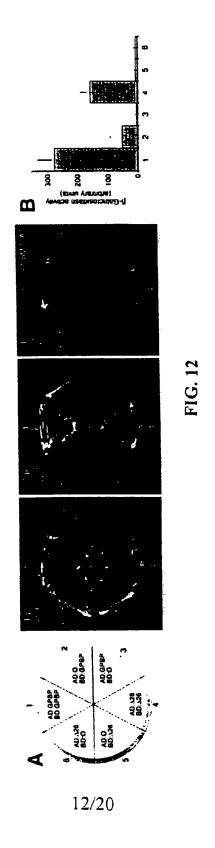


FIG. 11

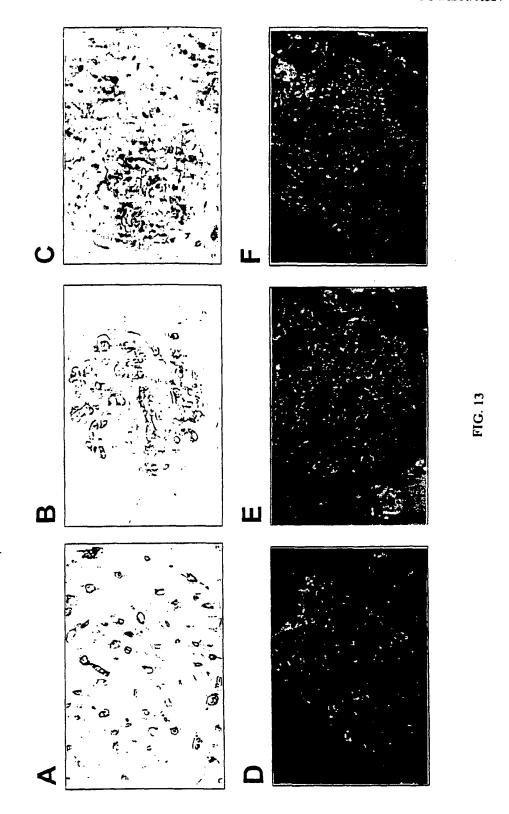
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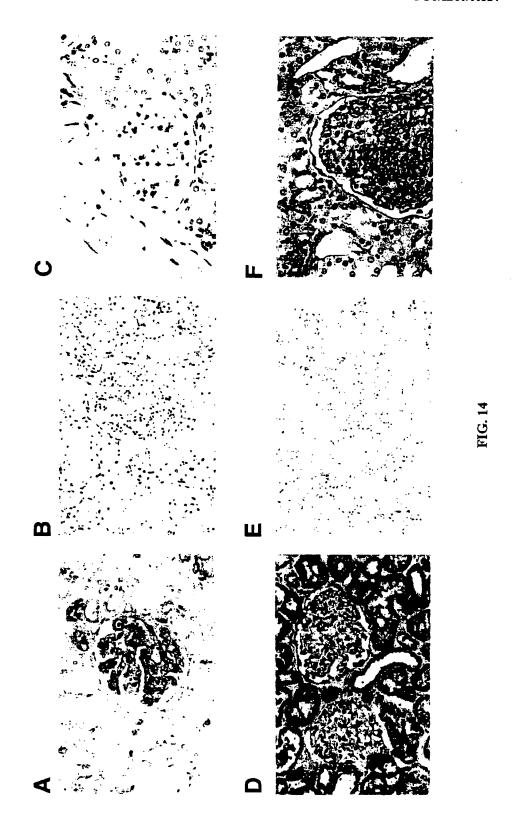
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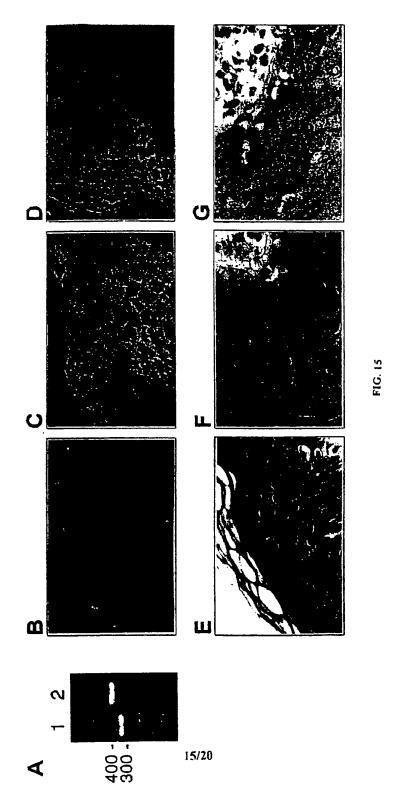
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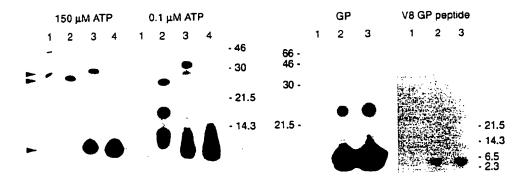


FIG. 16

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GPΔIII GLKGKRGDSGSPATWTTRGFVFTRHSQTTAI

MBP MASQKRP-SQRHGSKYLATASTMDHARHGFL

GPΔIII PSCPEGPVPLYSGFSFLFVQGNQRAHGQDLD

MBP PRHRDTGILDSIGRFFGGDRGAPKRGSGK-
GPΔIII ALFVKVLRSP

....|..||

MBP VPWLKPGRSP

FIG. 17

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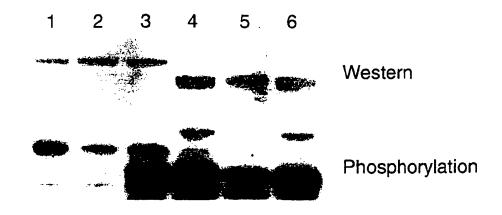


FIG. 18

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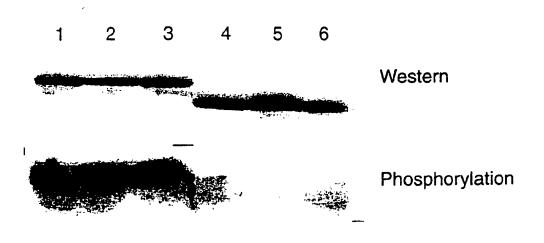


FIG. 19

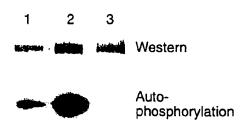


FIG. 20

20/20 SUBSTITUTE SHEET (RULE 26)

SEQUENCE LISTING

<110> Saus, Juan <120> Goodpasture Binding Protein <130> 98-723-B <140> To Be Assigned <141> Filed Herewith <160> 54 <170> PatentIn Ver. 2.0 <210> 1 <211> 2389 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (409)..(2280) gcaggaagat ggcggcggta gcggaggtgt gagtggacgc gggactcaqc ggccggattt 60 tetetteeet tettteeet ttteetteee tatttgaaat tggeategag ggggetaagt 120 tcgggtggca gcgccgggcg caacgcaggg gtcacggcga cggcggcggc ggctgacggc 180 tggaagggta ggcttcattc accgctcgtc ctccttcctc gctccgctcg gtgtcaggcg 240 cggcggcggc gcggcgggcg gacttcgtcc ctcctcctgc tccccccac accggagcgg 300 geactetteg ettegecate eccegaecet teacceegag gaetgggege etecteegge 360 gcagctgagg gagcgggggc cggtctcctg ctcggttgtc gagcctcc atg tcg gat 417 Met Ser Asp aat cag age tgg aac teg teg gge teg gag gag gat eea gag acg gag 465 Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu Thr Glu tot ggg cog cot gtg gag ogc tgc ggg gtc ctc agt aag tgg aca aac Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn 25 tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat aat gct 561 Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala 45 ctg agt tac tac aaa tct gaa gat gaa aca gag tat ggc tgc aga gga 609 Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly 55 tee ate tgt ett age aag get gte ate aca eet eac gat ttt gat gaa Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu 70

1

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tca Ser	atg Met	gtg Val	tcc Ser 135	ctg Leu	gtg Val	tct Ser	gga Gly	gca Ala 140	agt Ser	ggc	tac Tyr	tct Ser	gca Ala 145	aca Thr	tcc Ser	849
acc Thr	tct Ser	tca Ser 150	ttc Phe	aag Lys	aaa Lys	ggc Gly	cac His 155	agt Ser	tta Leu	cgt Arg	gag Glu	aag Lys 160	ttg Leu	gct Ala	gaa Glu	897
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gat	gct	gtt	gaa	gct	gct	ctt	gac	aga	caa	gat	aaa	ata	gaa	gaa	cag	1425

Asp	Ala 325	Val	Glu	Ala	Ala	Leu 330	Asp	Arg	Gln	Asp	Lys 335	Ile	Glu	Glu	Gln	
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ttg Leu 420	gtt Val	gta Val	gaa Glu	gaa Glu	gga Gly 425	gaa Glu	atg Met	aag Lys	gta Val	tac Tyr 430	aga Arg	aga Arg	gaa Glu	gta Val	gaa Glu 435	1713
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tta Leu	gct Ala 485	gat Asp	aat Asn	gca Ala	atc Ile	atc Ile 490	att Ile	tat Tyr	caa Gln	aca Thr	cac His 495	aag Lys	agg Arg	gtg Val	tgg Trp	1905
cct Pro 500	gct Ala	tct Ser	cag Gln	cga Arg	gac Asp 505	gta Val	tta Leu	tat Tyr	ctt Leu	tct Ser 510	gtc Val	att Ile	cga Arg	aag Lys	ata Ile 515	1953
cca Pro	gcc Ala	ttg Leu	act Thr	gaa Glu 520	aat Asn	gac Asp	cct Pro	gaa Glu	act Thr 525	tgg Trp	ata Ile	gtt Val	tgt Cys	aat Asn 530	ttt Phe	2001
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aaa Lys	ata Ile	aat Asn 550	gtt Val	gct Ala	atg Met	att Ile	tgt Cya 555	caa Gln	acc Thr	ttg Leu	gta Val	agc Ser 560	cca Pro	cca Pro	gag Glu	2097
gga Gly	aac Asn	cag Gln	gaa Glu	att Ile	agc Ser	agg Arg	gac Asp	aac Asn	att Ile	cta Leu	tgc Cys	aag Lys	att Ile	aca Thr	tat Tyr	2145

2290

2389

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Thr Leu Gln Lys Tyr Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp

Leu Ala Glu Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp

Glu Leu Gln Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro 195 200 205

- Thr Thr Arg Ser Asp Gly Asp Phe Leu His Ser Thr Asn Gly Asn Lys 210 215 220
- Glu Lys Leu Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp 225 230 235 240
- Phe Lys Gly Glu Ala Ile Thr Phe Lys Ala Thr Thr Ala Gly Ile Leu 245 250 255
- Ala Thr Leu Ser His Cys Ile Glu Leu Met Val Lys Arg Glu Asp Ser 260 265 270
- Trp Gln Lys Arg Leu Asp Lys Glu Thr Glu Lys Lys Arg Arg Thr Glu 275 280 285
- Glu Ala Tyr Lys Asn Ala Met Thr Glu Leu Lys Lys Ser His Phe 290 295 300
- Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu Ile Asn Glu Glu 305 310 315 320
- Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile 325 330 335
- Glu Glu Gln Ser Gln Ser Glu Lys Val Arg Leu His Trp Pro Thr Ser 340 345 350
- Leu Pro Ser Gly Asp Ala Phe Ser Ser Val Gly Thr His Arg Phe Val 355 360 365
- Gln Lys Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val 370 380
- Ser Ala Ser Asp Asp Val His Arg Phe Ser Ser Gln Val Glu Glu Met 385 390 395 400
- Val Gln Asn His Met Thr Tyr Ser Leu Gln Asp Val Gly Gly Asp Ala 405 410 415
- Asn Trp Gln Leu Val Val Glu Glu Glu Glu Met Lys Val Tyr Arg Arg
 420 425 430
- Glu Val Glu Asn Gly Ile Val Leu Asp Pro Leu Lys Ala Thr His 435 440 445
- Ala Val Lys Gly Val Thr Gly His Glu Val Cys Asn Tyr Phe Trp Asn 450 460
- Val Asp Val Arg Asn Asp Trp Glu Thr Thr Ile Glu Asn Phe His Val 465 470 475 480
- Val Glu Thr Leu Ala Asp Asn Ala Ile Ile Ile Tyr Gln Thr His Lys 485 490 495
- Arg Val Trp Pro Ala Ser Gln Arg Asp Val Leu Tyr Leu Ser Val Ile 500 505 510
- Arg Lys Ile Pro Ala Leu Thr Glu Asn Asp Pro Glu Thr Trp Ile Val

515 520 525

Cys Asn Phe Ser Val Asp His Asp Ser Ala Pro Leu Asn Asn Arg Cys 530 540

Val Arg Ala Lys Ile Asn Val Ala Met Ile Cys Gln Thr Leu Val Ser 545 550 555 560

Pro Pro Glu Gly Asn Gln Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys 565 570 575

Ile Thr Tyr Val Ala Asn Val Asn Pro Gly Gly Trp Ala Pro Ala Ser 580 585 590

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Met Ser Asp Asn Gln Ser Trp Asn Ser Ser

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tgc ggg gtc ctc agc aag tgg aca aac tat att cat gga tgg cag gat 569 Cys Gly Val Leu Ser Lys Trp Thr Asn Tyr Ile His Gly Trp Gln Asp 30 35 40

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	atc Ile															713
	gat Asp															761
	tgg Trp															809
	gag Glu															857
	gcg Ala 140															905
	agt Ser															953
	tgc Cys															1001
	gct Ala															1049
	gaa Glu															1097
	acc Thr 220															1145
	att Ile					Phe					Ile					1193
	act Thr				Leu					His						1241
				Glu					Arg					Val	gaa Glu	1289
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aag	aag	aaa	ccc	: cgt	tto	gga	ggg	cce	gat	tat	gaa	gaa	ggt	cca	aac	1385

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Gly	Thr	His 365	Arg	Phe	Val	Gln	Lys 370	Pro	Tyr	Ser	Arg	Ser 375	tcc Ser	Ser	Met	1577
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Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly
575 580 585

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Asn Asn Thr Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly 50 55 60

Cys Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp 65 70 75 80

Phe Asp Glu Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr
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Leu Arg Ala Gln Asp Pro Glu His Arg Gln Gln Trp Val Asp Ala Ile 100 105 110

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- Arg His Gly Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser 130 140
- Ala Thr Ser Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys 145 150 155 160
- Leu Ala Glu Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp 165 170 175
- Thr Leu Gln Lys Tyr Phe Asp Val Cys Ala Asp Ala Val Ser Lys Asp 180 185 190
- Glu Leu Gln Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro 195 200 205
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- Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile 325 330 335
- Glu Glu Gln Ser Gln Ser Glu Lys Val Arg Leu His Trp Pro Thr Ser 340 345 350
- Leu Pro Ser Gly Asp Thr Phe Ser Ser Val Gly Thr His Arg Phe Val 355 360 365
- Gln Lys Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val
- Ser Ala Ser Asp Asp Val His Arg Phe Ser Ser Gln Val Glu Glu Met 385 390 395 400
- Val Gln Asn His Met Asn Tyr Ser Leu Gln Asp Val Gly Gly Asp Ala
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- Asn Trp Gln Leu Val Val Glu Glu Gly Glu Met Lys Val Tyr Arg Arg

420 425 430

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Ala Val Lys Gly Val Thr Gly His Glu Val Cys Asn Tyr Phe Trp Asn 450 460

Val Asp Val Arg Asn Asp Trp Glu Thr Thr Ile Glu Asn Phe His Val 465 470 475 480

Val Glu Thr Leu Ala Asp Asn Ala Ile Ile Val Tyr Gln Thr His Lys 485 490 495

Arg Val Trp Pro Ala Ser Gln Arg Asp Val Leu Tyr Leu Ser Ala Ile 500 505 510

Arg Lys Ile Pro Ala Leu Thr Glu Asn Asp Pro Glu Thr Trp Ile Val 515 520 525

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_		Arg		_		_	Phe	_				Asn		aat Asn	_	1092
	Lys					Val					Ile			ata Ile		1140

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- Arg His Gly Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser 130 140
- Ala Thr Ser Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys 145 150 155 160
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- Gln Lys Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val 370 380
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Val Gln Asn His Met Thr Tyr Ser Leu Gln Asp Val Gly Gly Asp Ala 405 410 415

Asn Trp Gln Leu Val Val Glu Glu Gly Glu Met Lys Val Tyr Arg Arg 420 425 430

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Arg Lys Ile Pro Ala Leu Asn Glu Asn Asp Pro Glu Thr Trp Ile Val 515 520 525

Cys Asn Phe Ser Val Asp His Ser Ser Ala Pro Leu Asn Asn Arg Cys 530 540

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Leu Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile 100 105 110

Glu Gln His Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg 115 120 125

- Arg His Gly Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser 130 140
- Ala Thr Ser Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys 145 150 155 160
- Leu Ala Glu Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp 165 170 175
- Thr Leu Gln Lys Tyr Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp 180 185 190
- Glu Leu Gln Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro 195 200 205
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- Glu Lys Leu Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp 225 230 235 240
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- Trp Gln Lys Arg Leu Asp Lys Glu Thr Glu Lys Lys Arg Arg Thr Glu 275 280 285
- Glu Ala Tyr Lys Asn Ala Met Thr Glu Leu Lys Lys Lys Ser His Phe 290 295 300
- Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu Ile Asn Glu Glu 305 310 315 320
- Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile
- Glu Glu Gln Ser Gln Ser Glu Lys Val Arg Leu His Trp Pro Thr Ser 340 345
- Leu Pro Ser Gly Asp Ala Phe Ser Ser Val Gly Thr His Arg Phe Val 355 360 365
- Gln Lys Val Glu Glu Met Val Gln Asn His Met Thr Tyr Ser Leu Gln 370 $$ 380
- Asp Val Gly Gly Asp Ala Asn Trp Gln Leu Val Val Glu Glu Gly Glu 385 390 395
- Met Lys Val Tyr Arg Arg Glu Val Glu Glu Asn Gly Ile Val Leu Asp 405 410 415
- Pro Leu Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His Glu Val
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Ile Glu Asn Phe His Val Val Glu Thr Leu Ala Asp Asn Ala Ile Ile

Ile Tyr Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg Asp Val

Leu Tyr Leu Ser Val Ile Arg Lys Ile Pro Ala Leu Thr Glu Asn Asp

Pro Glu Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Asp Ser Ala

Pro Leu Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Val Ala Met Ile

Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asn Gln Glu Ile Ser Arg 535

Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly 555 545

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Gln Lys Val Glu Glu Met Val Gln Asn His Met Asn Tyr Ser Leu Gln

375

380

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Pro Leu Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His Glu Val 420 425 430

Cys Asn Tyr Phe Trp Asn Val Asp Val Arg Asn Asp Trp Glu Thr Thr 435 440 445

Ile Glu Asn Phe His Val Val Glu Thr Leu Ala Asp Asn Ala Ile Ile 450 460

Val Tyr Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg Asp Val 465 470 475 480

Leu Tyr Leu Ser Ala Ile Arg Lys Ile Pro Ala Leu Thr Glu Asn Asp 485 490 495

Pro Glu Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Asp Ser Ala 500 505 510

Pro Leu Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Ile Ala Met Ile 515 520 525

Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asp Gln Glu Ile Ser Arg 530 535 540

Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly 545 550 555 560

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gct aca ct Ala Thr Le	t tct cat u Ser His 260	tgt att Cys Ile	gag ctg Glu Leu 265	Met V	ta aaa al Lys	cgt ga Arg Gl 27	u Asp	agc Ser	1236
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gag ttc tt Glu Phe Ph									1428
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atg cca to Met Pro Se 35	r Gly Asp	-				_		-	1524
caa aag gt Gln Lys Va 370			Gln Ası						1572
gat gta gg Asp Val Gl 385				Leu V					1620
atg aag gt Met Lys Va		Arg Glu			Asn Gly				1668
cct ttg aa Pro Leu Ly	a gct acc /s Ala Thr 420	cat gca His Ala	gtt aa: Val Ly:	Gly V	gtt aca Val Thr	gga ca Gly Hi 43	s Glu	gtc Val	1716
tgc aat ta Cys Asn Ty 4:									1764
ata gaa aa Ile Glu As 450			Glu Th						1812

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363 390	
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29

85 90 9

Leu Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile
100 105 110

Glu Gln His Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg 115 120 125

Arg His Gly Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser

Ala Thr Ser Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys 145 150 155 160

Leu Ala Glu Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp
165 170 175

Thr Leu Gln Lys Phe Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp 180 185 190

Glu Phe Gln Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro 195 200 205

Thr Thr Arg Ser Asp Gly Asp Phe Leu His Asn Thr Asn Gly Asn Lys 210 215 220

Glu Lys Val Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp 225 230 235 240

Phe Lys Gly Glu Ala Ile Thr Phe Lys Ala Thr Thr Ala Gly Ile Leu 245 250 255

Ala Thr Leu Ser His Cys Ile Glu Leu Met Val Lys Arg Glu Asp Ser 260 265 270

Trp Gln Lys Arg Met Asp Lys Glu Thr Glu Lys Arg Arg Val Glu 275 280 285

Glu Ala Tyr Lys Asn Ala Met Thr Glu Leu Lys Lys Lys Ser His Phe 290 295 300

Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu Ile Asn Glu Glu 305 310 315

Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile 325 330 335

Glu Glu Gln Ser Gln Ser Glu Lys Val Arg Leu His Trp Ser Thr Ser 340 345 350

Met Pro Ser Gly Asp Ala Phe Ser Ser Val Gly Thr His Arg Phe Val 355 360 365

Gln Lys Val Glu Glu Met Val Gln Asn His Met Thr Tyr Ser Leu Gln 370 375 380

Asp Val Gly Gly Asp Ala Asn Trp Gln Leu Val Val Glu Glu Gly Glu 385 390 395

Met Lys Val Tyr Arg Arg Glu Val Glu Glu Asn Gly Ile Val Leu Asp 405 410 415

Pro Leu Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His Glu Val Cys Asn Tyr Phe Trp Asn Val Asp Val Arg Asn Asp Trp Glu Thr Thr 440 Ile Glu Asn Phe His Val Val Glu Thr Leu Ala Asp Asn Ala Ile Ile 455 Ile Tyr Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg Asp Val 470 Leu Tyr Leu Ser Ala Ile Arg Lys Ile Pro Ala Leu Asn Glu Asn Asp Pro Glu Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Ser Ser Ala Pro Leu Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Val Ala Met Ile Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asn Gln Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg Ala Val Ala Lys Arg Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln Glu Lys Thr Ala Gly Lys Pro Ile Leu Phe 595 <210> 13 <211> 78 <212> DNA <213> Homo sapiens <220> <221> CDS <222> (1)..(78) <400> 13 ecc tat agt ege tet tee tee atg tet tee att gat eta gte agt gee Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val Ser Ala 10 tot gat gat gtt cac aga ttc agc tcc cag 78 Ser Asp Asp Val His Arg Phe Ser Ser Gln 20

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Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val Ser Ala Ser Asp Asp Val His Arg Phe Ser Ser Gln 20 <210> 15 <211> 2034 <212> DNA <213> Artificial Sequence <223> Description of Artificial Sequence: GPBPR3 <220> <221> CDS <222> (10) .. (990) <400> 15 gaattcacc atg gcc cca cta gcc gac tac aag gac gac gat gac aag atg 51 Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Asp Lys Met tog gat aat cag ago tgg aac tog tog ggc tog gag gat coa gag 99 Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu acg gag tot ggg cog cot gtg gag ogo tgc ggg gto oto agt aag tgg 147 Thr Glu Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp 35 aca aac tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat 195 Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn 55 aat gct ctg agt tac tac aaa tct gaa gat gaa aca gag tat ggc tgc 243 Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys aga gga tcc atc tgt ctt agc aag gct gtc atc aca cct cac gat ttt 291 Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe 80 gat gaa tgt cga ttt gat att agt gta aat gat agt gtt tgg tat ctt 339 Asp Glu Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu 95 100 cgt gct cag gat cca gat cat aga cag caa tgg ata gat gcc att gaa 387 Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu 115 120 cag cac aag act gaa tot gga tat gga tot gaa too ago ttg cgt cga 435 Gln His Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg cat ggc tca atg gtg tcc ctg gtg tct gga gca agt ggc tac tct gca His Gly Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser Ala 150

	tcc Ser 160											_		_	-	531
	gaa Glu															579
	cag Gln	_			_		_	_	-	_	_		_	_	-	627
	caa Gln															675
	cgt Arg															723
_	tta Leu 240													_		771
	gly aaa															819
	ctt L eu															867
	aag Lys															915
	tat Tyr															963
-	att Ile 320	_	_	_	_			-	_	ttaa	tga	agaa	gagt	tc		1010
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<212> PRT

<213> Artificial Sequence

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<223> Description of Artificial Sequence: GPBPR3

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Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn 35 40 45

Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala 50 55 60

Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly 65 70 75 80

Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu 85 90 95

Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu Arg Ala

Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu Gln His 115 120 125

Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg His Gly
130 135 140

Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser Ala Thr Ser 145 150 155 160

Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys Leu Ala Glu

165 170 175

Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp Thr Leu Gln
180 185 190

Lys Tyr Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp Glu Leu Gln 195 200 205

Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro Thr Thr Arg 210 215 220

Ser Asp Gly Asp Phe Leu His Ser Thr Asn Gly Asn Lys Glu Lys Leu 235 240

Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp Phe Lys Gly 245 250 255

Glu Ala Ile Thr Phe Lys Ala Thr Thr Ala Gly Ile Leu Ala Thr Leu 260 265 270

Ser His Cys Ile Glu Leu Met Val Lys Arg Glu Asp Ser Trp Gln Lys 275 280 285

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<211> 1978

<212> DNA

<213> Artificial Sequence

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<222> (10)..(1860)

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Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu
15 20 25 30

acg gag tct ggg ccg cct gtg gag cgc tgc ggg gtc ctc agt aag tgg 147
Thr Glu Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp
35 40 45

aca aac tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat 195
Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn
50 55

aat Asn	gct Ala	ctg Leu 65	agt Ser	tac Tyr	tac Tyr	aaa Lys	tct Ser 70	gaa Glu	gat Asp	gaa Glu	aca Thr	gag Glu 75	tat Tyr	ggc Gly	tgc Cys	243
aga Arg	gga Gly 80	tcc Ser	atc Ile	tgt Cys	ctt Leu	agc Ser 85	aag Lys	gct Ala	gtc Val	atc Ile	aca Thr 90	cct Pro	cac His	gat Asp	ttt Phe	291
gat Asp 95	gaa Glu	tgt Cys	cga Arg	ttt Phe	gat Asp 100	att Ile	agt Ser	gta Val	aat Asn	gat Asp 105	agt Ser	gtt Val	tgg Trp	tat Tyr	ctt Leu 110	339
cgt Arg	gct Ala	cag Gln	gat Asp	cca Pro 115	gat Asp	cat His	aga Arg	cag Gln	caa Gln 120	tgg Trp	ata Ile	gat Asp	gcc Ala	att Ile 125	gaa Glu	387
cag Gln	cac His	aag Lys	act Thr 130	gaa Glu	tct Ser	gga Gly	tat Tyr	gga Gly 135	tct Ser	gaa Glu	tcc Ser	agc Ser	ttg Leu 140	cgt Arg	cga Arg	435
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aca Thr	tcc Ser 160	acc Thr	tct Ser	tca Ser	ttc Phe	aag Lys 165	aaa Lys	ggc Gly	cac His	agt Ser	tta Leu 170	cgt Arg	gag Glu	aag Lys	ttg Leu	531
gct Ala 175	gaa Glu	atg Met	gaa Glu	aca Thr	ttt Phe 180	aga Arg	gac Asp	atc Ile	tta Leu	tgt Cys 185	aga Arg	caa Gln	gtt Val	gac Asp	acg Thr 190	579
cta Leu	cag Gln	aag Lys	tac Tyr	ttt Phe 195	gat Asp	gcc Ala	tgt Cys	gct Ala	gat Asp 200	gct Ala	gtc Val	tct Ser	aag Lys	gat Asp 205	gaa Glu	627
ctt Leu	caa Gln	agg Arg	gat Asp 210	aaa Lys	gtg Val	gta Val	gaa Glu	gat Asp 215	gat Asp	gaa Glu	gat Asp	gac Asp	ttt Phe 220	cct Pro	aca Thr	675
acg Thr	cgt Arg	tct Ser 225	gat Asp	ggt Gly	gac Asp	ttc Phe	ttg Leu 230	cat His	agt Ser	acc Thr	aac Asn	ggc Gly 235	aat Asn	aaa Lys	gaa Glu	723
aag Lys	tta Leu 240	ttt Phe	cca Pro	cat His	gtg Val	aca Thr 245	cca Pro	aaa Lys	gga Gly	att Ile	aat Asn 250	ggt Gly	ata Ile	gac Asp	ttt Phe	771
aaa Lys 255	61 y 999	gaa Glu	gcg Ala	ata Ile	act Thr 260	ttt Phe	aaa Lys	gca Ala	act Thr	act Thr 265	gct Ala	gga Gly	atc Ile	ctt Leu	gca Ala 270	819
aca Thr	ctt Leu	tct Ser	cat His	tgt Cys 275	att Ile	g aa Glu	cta Leu	atg Met	gtt Val 280	aaa Lys	cgt Arg	gag Glu	gac Asp	agc Ser 285	tgg Trp	867
cag Gln	aag Lys	aga Arg	ctg Leu 290	gat Asp	aag Lys	gaa Glu	act Thr	gag Glu 295	cac His	ttt Phe	gga Gly	gga Gly	cca Pro 300	gat Asp	tat Tyr	915

gaa Glu	gaa Glu	ggc Gly 305	cct Pro	aac Asn	agt Ser	ctg Leu	att Ile 310	aat Asn	g aa Glu	gaa Glu	gag Glu	ttc Phe 315	ttt Phe	gat Asp	gct Ala	963
gtt Val	gaa Glu 320	gct Ala	gct Ala	ctt Leu	gac Asp	aga Arg 325	caa Gln	gat Asp	aaa Lys	ata Ile	gaa Glu 330	gaa Glu	cag Gln	tca Ser	cag Gln	1011
agt Ser 335	gaa Glu	aag Lys	gtg Val	aga Arg	tta Leu 340	cat His	tgg Trp	cct Pro	aca Thr	tcc Ser 345	ttg Leu	ccc Pro	tct Ser	gga Gly	gat Asp 350	1059
gcc Ala	ttt Phe	tct Ser	tct Ser	gtg Val 355	ggg Gly	aca Thr	cat His	aga Arg	ttt Phe 360	gtc Val	caa Gln	aag Lys	ccc Pro	tat Tyr 365	agt S e r	1107
Arg	Ser	Ser	Ser 370	Met	Ser	Ser	Ile	Asp 375	Leu	Val	Ser	Ala	Ser 380	gat Asp	Asp	1155
Val	His	Arg 385	Phe	Ser	Ser	Gln	Val 390	Glu	Glu	Met	Val	Gln 395	Asn	cac His	Met	1203
Thr	Tyr 400	Ser	Leu	Gln	Asp	Val 405	Gly	Gly	Asp	Ala	Asn 410	Trp	Gln	ttg Leu	Val	1251
Val 415	Glu	Glu	Gly	Glu	Met 420	ГÀа	Val	Tyr	Arg	Arg 425	Glu	Val	Glu	gaa Glu	Asn 430	1299
Gly	Ile	Val	Leu	Asp 435	Pro	Leu	Lys	Ala	Thr 440	His	Ala	Val	Lys	ggc Gly 445	Val	1347
Thr	Gly	His	Glu 450	Val	Сув	Asn	Tyr	Phe 455	Trp	Asn	Val	Asp	Val 460	cgc Arg	Asn	1395
Asp	Trp	Glu 465	Thr	Thr	Ile	Glu	Asn 470	Phe	His	Val	Val	Glu 475	Thr	tta Leu	Ala	1443
Asp	Asn 480	Ala	Ile	Ile	Ile	Tyr 485	Gln	Thr	His	ГÀЗ	Arg 490	Val	Trp	cct Pro	Ala	1491
Ser 495	Gln	Arg	Asp	Val	Leu 500	Tyr	Leu	Ser	Val	Ile 505	Arg	Lys	Ile	cca Pro	Ala 510	1539
Leu	Thr	Glu	Asn	Asp 515	Pro	Glu	Thr	Trp	11e 520	Val	Cys	Asn	Phe	tct Ser 525	Val	1587
Asp	His	Asp	Ser 530	Ala	Pro	Leu	Asn	Asn 535	Arg	Cys	Val	Arg	Ala 540	aaa Lys	Ile	1635
aat	gtt	gct	atg	att	tgt	caa	acc	ttg	gta	agc	cca	cca	gag	gga	aac	1683

Asn Val Ala Met Ile Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asn 545 550 cag gaa att agc agg gac aac att cta tgc aag att aca tat gta gct 1731 Gln Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala aat gtg aac cct gga gga tgg gca cca gcc tca gtg tta agg gca gtg Asn Val Asn Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg Ala Val 580 gca aag cga gag tat cct aaa ttt cta aaa cgt ttt act tct tac gtc Ala Lys Arg Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val 595 600 caa gaa aaa act gca gga aag cct att ttg ttc tagtattaac aggtactaga 1880 Gln Glu Lys Thr Ala Gly Lys Pro Ile Leu Phe agatatgttt tatctttttt taactttatt tgactaatat gactgtcaat actaaaattt 1940 agttgttgaa agtatttact atgttttttc cggaattc <210> 18 <211> 617 <212> PRT <213> Artificial Sequence <220> <223> Description of Artificial Sequence: FLAG-GPBPDNLS Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu Thr Glu 25 Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu Gln His Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg His Gly

135

Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser Ala Thr Ser

150 155 Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys Leu Ala Glu Met Glu Thr Phe Arg Asp Ile Leu Cys Arg Gln Val Asp Thr Leu Gln Lys Tyr Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp Glu Leu Gln Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro Thr Thr Arg Ser Asp Gly Asp Phe Leu His Ser Thr Asn Gly Asn Lys Glu Lys Leu Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp Phe Lys Gly Glu Ala Ile Thr Phe Lys Ala Thr Thr Ala Gly Ile Leu Ala Thr Leu 265 Ser His Cys Ile Glu Leu Met Val Lys Arg Glu Asp Ser Trp Gln Lys 280 Arg Leu Asp Lys Glu Thr Glu His Phe Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu Ile Asn Glu Glu Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile Glu Glu Gln Ser Gln Ser Glu 325 330 Lys Val Arg Leu His Trp Pro Thr Ser Leu Pro Ser Gly Asp Ala Phe 345 Ser Ser Val Gly Thr His Arg Phe Val Gln Lys Pro Tyr Ser Arg Ser 360 Ser Ser Met Ser Ser Ile Asp Leu Val Ser Ala Ser Asp Asp Val His Arg Phe Ser Ser Gln Val Glu Glu Met Val Gln Asn His Met Thr Tyr 390 Ser Leu Gln Asp Val Gly Gly Asp Ala Asn Trp Gln Leu Val Val Glu 405 Glu Gly Glu Met Lys Val Tyr Arg Arg Glu Val Glu Glu Asn Gly Ile 425 Val Leu Asp Pro Leu Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His Glu Val Cys Asn Tyr Phe Trp Asn Val Asp Val Arg Asn Asp Trp 455 Glu Thr Thr Ile Glu Asn Phe His Val Val Glu Thr Leu Ala Asp Asn

Ala Ile Ile Ile Tyr Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg Asp Val Leu Tyr Leu Ser Val Ile Arg Lys Ile Pro Ala Leu Thr 505 Glu Asn Asp Pro Glu Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Asp Ser Ala Pro Leu Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Val Ala Met Ile Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asn Gln Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg Ala Val Ala Lys 580 585 Arg Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln Glu 600 Lys Thr Ala Gly Lys Pro Ile Leu Phe <210> 19 <211> 1975 <212> DNA <213> Artificial Sequence <220> <223> Description of Artificial Sequence: FLAG-GPBPDSXY <220> <221> CDS <222> (10) .. (1857) <400> 19 gaattcacc atg gcc cca cta gcc gac tac aag gac gac gat gac aag atg 51 Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met tog gat aat cag ago tgg aac tog tog ggc tog gag gag gat coa gag Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu acg gag tot ggg cog cot gtg gag ogc tgc ggg gtc otc agt aag tgg 147 Thr Glu Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp 40 aca aac tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn aat get etg agt tae tae aaa tet gaa gat gaa aca gag tat gge tge 243

Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys

70

65

aga Arg	gga Gly 80	tcc Ser	atc Ile	tgt Cys	ctt Leu	agc Ser 85	aag Lys	gct Ala	gtc Val	atc Ile	aca Thr 90	cct Pro	cac His	gat Asp	ttt Phe	291
gat Asp 95	gaa Glu	tgt Cys	cga Arg	ttt Phe	gat Asp 100	att Ile	agt Ser	gta Val	aat Asn	gat Asp 105	agt Ser	gtt Val	tgg Trp	tat Tyr	ctt Leu 110	339
cgt Arg	gct Ala	cag Gln	gat Asp	cca Pro 115	gat Asp	cat His	aga Arg	cag Gln	caa Gln 120	tgg Trp	ata Ile	gat Asp	gcc Ala	att Ile 125	gaa Glu	387
Gln	His	Lys	Thr 130	Glu	tct Ser	Gly	Tyr	Gly 135	Ser	Glu	Ser	Ser	Leu 140	Arg	Arg	435
His	Gly	Lys 145	Gly	His	agt Ser	Leu	Arg 150	Glu	Lys	Leu	Ala	Glu 155	Met	Glu	Thr	483
Phe	Arg 160	Asp	Ile	Leu	tgt Cys	Arg 165	Gln	Val	Asp	Thr	Leu 170	Gln	Lys	Tyr	Phe	531
Asp 175	Ala	Cys	Ala	Asp	gct Ala 180	Val	Ser	Lys	qaA	Glu 185	Leu	Gln	Arg	Asp	Lys 190	579
gtg Val	gta Val	gaa Glu	gat Asp	gat Asp 195	gaa Glu	gat Asp	gac Asp	ttt Phe	cct Pro 200	aca Thr	acg Thr	cgt Arg	tct Ser	gat Asp 205	ggt Gly	627
qaA	Phe	Leu	His 210	Ser	acc Thr	Asn	Gly	Asn 215	Lys	Glu	Lys	Leu	Phe 220	Pro	His	675
Val	Thr	Pro 225	Lys	Gly	att Ile	Asn	Gly 230	Ile	Asp	Phe	Lys	Gly 235	Glu	Ala	Ile	723
Thr	Phe 240	Lys	Ala	Thr	act Thr	Ala 245	Gly	Ile	Leu	Ala	Thr 250	Leu	Ser	His	Сув	771
11e 255	Glu	Leu	Met	Val	aaa Lys 260	Arg	Glu	qsA	Ser	Trp 265	Gln	Lys	Arg	Leu	Asp 270	819
aag Lys	gaa Glu	act Thr	gag Glu	aag Lys 275	aaa Lys	aga Arg	aga Arg	aca Thr	gag Glu 280	gaa Glu	gca Ala	tat Tyr	aaa Lys	aat Asn 285	gca Ala	867
Met	Thr	Glu	Leu 290	Lys	aaa Lys	Lys	Ser	His 295	Phe	Gly	Gly	Pro	Asp 300	Tyr	Glu	915
gaa Glu	ggc Gly	CCT Pro 305	aac Asn	agt Ser	ctg Leu	att Ile	aat Asn 310	gaa Glu	gaa Glu	gag Glu	ttc Phe	ttt Phe 315	gat Asp	gct Ala	gtt Val	963

	gct Ala 320															1011
	aag Lys															1059
	tct Ser															1107
	tcc Ser															1155
	aga Arg															1203
	tca Ser 400															1251
	gaa Glu															1299
	gtt Val	_	_				-			_	_			_		1347
	cat His															1395
	gaa Glu															1443
	gca Ala 480															1491
	cga Arg		Val		Tyr	Leu		Val	Ile	Arg	Lys		Pro			1539
Thr	Glu	Asn	Asp	Pro 515	Glu	Thr	Trp	Ile	Val 520	Сув	Asn	Phe	Ser	Val 525		1587
	gac Asp													Ile	aat Asn	1635
	_		Ile	_			_	Val					~ ~		cag Gln	1683
gaa	att	ago	agg	gac	aac	att	cta	tgo	aag	att	aca	tat	gta	gct	aat	1731

Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn 565 gtg aac cot gga gga tgg gca coa gco toa gtg tta agg gca gtg gca Val Asn Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg Ala Val Ala 580 585 aag cga gag tat cet aaa ttt cta aaa cgt ttt act tet tae gte caa Lys Arg Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln gaa aaa act gca gga aag cct att ttg ttc tagtattaac aggtactaga 1877 Glu Lys Thr Ala Gly Lys Pro Ile Leu Phe 610 agatatgttt tatctttttt taactttatt tgactaatat gactgtcaat actaaaattt 1937 agttgttgaa agtatttact atgttttttc cggaattc 1975 <210> 20 <211> 616 <212> PRT <213> Artificial Sequence <223> Description of Artificial Sequence: FLAG-GPBPDSXY <400> 20 Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu Thr Glu 20 Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu Gln His Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg His Gly 135 Lys Gly His Ser Leu Arg Glu Lys Leu Ala Glu Met Glu Thr Phe Arg

170

Asp Ile Leu Cys Arg Gln Val Asp Thr Leu Gln Lys Tyr Phe Asp Ala

165

Cys Ala Asp Ala Val Ser Lys Asp Glu Leu Gln Arg Asp Lys Val Val 180 185 190

- Glu Asp Asp Glu Asp Asp Phe Pro Thr Thr Arg Ser Asp Gly Asp Phe
 195 200 205
- Leu His Ser Thr Asn Gly Asn Lys Glu Lys Leu Phe Pro His Val Thr 210 215 220
- Pro Lys Gly Ile Asn Gly Ile Asp Phe Lys Gly Glu Ala Ile Thr Phe 225 230 235 240
- Lys Ala Thr Thr Ala Gly Ile Leu Ala Thr Leu Ser His Cys Ile Glu 245 250 255
- Leu Met Val Lys Arg Glu Asp Ser Trp Gln Lys Arg Leu Asp Lys Glu 260 265 270
- Thr Glu Lys Lys Arg Arg Thr Glu Glu Ala Tyr Lys Asn Ala Met Thr
- Glu Leu Lys Lys Lys Ser His Phe Gly Gly Pro Asp Tyr Glu Glu Gly 290 295 300
- Pro Asn Ser Leu Ile Asn Glu Glu Glu Phe Phe Asp Ala Val Glu Ala 305 310 315 320
- Ala Leu Asp Arg Gln Asp Lys Ile Glu Glu Gln Ser Gln Ser Glu Lys 325 330 335
- Val Arg Leu His Trp Pro Thr Ser Leu Pro Ser Gly Asp Ala Phe Ser 340 345 350
- Ser Val Gly Thr His Arg Phe Val Gln Lys Pro Tyr Ser Arg Ser Ser 355 360 365
- Ser Met Ser Ser Ile Asp Leu Val Ser Ala Ser Asp Asp Val His Arg 370 375 380
- Phe Ser Ser Gln Val Glu Glu Met Val Gln Asn His Met Thr Tyr Ser 385 395 400
- Leu Gln Asp Val Gly Gly Asp Ala Asn Trp Gln Leu Val Val Glu Glu 415
- Gly Glu Met Lys Val Tyr Arg Arg Glu Val Glu Glu Asn Gly Ile Val 420 425 430
- Leu Asp Pro Leu Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His
 435 440 445
- Glu Val Cys Asn Tyr Phe Trp Asn Val Asp Val Arg Asn Asp Trp Glu 450 455 460
- Thr Thr Ile Glu Asn Phe His Val Val Glu Thr Leu Ala Asp Asn Ala 465 470 475 480
- Ile Ile Ile Tyr Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg 485 490 495

Asp Val Leu Tyr Leu Ser Val Ile Arg Lys Ile Pro Ala Leu Thr Glu 500 505 510

Asn Asp Pro Glu Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Asp 515 520 525

Ser Ala Pro Leu Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Val Ala 530 535 540

Met Ile Cys Gln Thr Leu Val Ser Pro Pro Glu Gly Asn Gln Glu Ile 545 550 555 560

Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn 565 570 575

Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg Ala Val Ala Lys Arg 580 585 590

Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln Glu Lys 595 600 605

Thr Ala Gly Lys Pro Ile Leu Phe 610 615

<210> 21

<211> 1915

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:
 FLAG-GPBPDSXY/NLS

(220>

<221> CDS

<222> (10)..(1797)

<400> 21

gaattcacc atg gcc cca cta gcc gac tac aag gac gac gat gac aag atg 51

Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met

1 5 10

tcg gat aat cag agc tgg aac tcg tcg ggc tcg gag gag gat cca gag 99
Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu
15 20 25

acg gag tct ggg ccg cct gtg gag cgc tgc ggg gtc ctc agt aag tgg 147 Thr Glu Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp 35 40

aca aac tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat 195
Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn
50 55

aat gct ctg agt tac tac aaa tct gaa gat gaa aca gag tat ggc tgc 243 Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys 65 70 75

aga gga tcc atc tgt ctt agc aag gct gtc atc aca cct cac gat ttt 291 Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe 80 85 90

					gat Asp 100											339
cgt Arg	gct Ala	cag Gln	gat Asp	cca Pro 115	gat Asp	cat His	aga Arg	cag Gln	caa Gln 120	tgg Trp	ata Ile	gat Asp	gcc Ala	att Ile 125	gaa Glu	387
cag Gln	cac His	aag Lys	act Thr 130	gaa Glu	tct Ser	gga Gly	tat Tyr	gga Gly 135	tct Ser	gaa Glu	tcc Ser	agc Ser	ttg Leu 140	cgt Arg	cga Arg	435
cat His	ggc Gly	aaa Lys 145	ggc Gly	cac His	agt Ser	tta Leu	cgt Arg 150	gag Glu	aag Lys	ttg Leu	gct Ala	gaa Glu 155	atg Met	gaa Glu	aca Thr	483
ttt Phe	aga Arg 160	gac Asp	atc Ile	tta Leu	tgt Cys	aga Arg 165	caa Gln	gtt Val	gac Asp	acg Thr	cta Leu 170	cag Gln	aag Lys	tac Tyr	ttt Phe	531
gat Asp 175	gcc Ala	tgt Cys	gct Ala	gat Asp	gct Ala 180	gtc Val	tct Ser	aag Lys	gat Asp	gaa Glu 185	ctt Leu	caa Gln	agg Arg	gat Asp	aaa Lys 190	579
					gaa Glu											627
					acc Thr											675
gtg Val	aca Thr	cca Pro 225	aaa Lys	gga Gly	att Ile	aat Asn	ggt Gly 230	ata Ile	gac Asp	ttt Phe	aaa Lys	999 Gly 235	gaa Glu	gcg Ala	ata Ile	723
act Thr	ttt Phe 240	aaa Lys	gca Ala	act Thr	act Thr	gct Ala 245	gga Gly	atc Ile	ctt Leu	gca Ala	aca Thr 250	ctt Leu	tct Ser	cat His	tgt Cys	771
att Ile 255	gaa Glu	cta Leu	atg M e t	gtt Val	aaa Lys 260	cgt Arg	gag Glu	gac Asp	agc Ser	tgg Trp 265	cag Gln	aag Lys	aga Arg	ctg Leu	gat Asp 270	819
					ttt Phe											867
					gaa Glu											915
g a c Asp	aga Arg	caa Gln 305	gat Asp	aaa Lys	ata Ile	gaa Glu	gaa Glu 310	cag Gln	tca Ser	cag Gln	agt Ser	gaa Glu 315	aag Lys	gtg Val	aga Arg	963
tta Leu	cat His 320	tgg Trp	cct Pro	aca Thr	tcc Ser	ttg Leu 325	ccc Pro	tct Ser	gga Gly	gat Asp	gcc Ala 330	ttt Phe	tct Ser	tct Ser	gtg Val	1011

			-		-		_			_	_	tct Ser			-	1059
												cac His				1107
	_	_	-				_			_		tac Tyr			_	1155
												gaa Glu 395				1203
												att Ile				1251
			_			_	-			-		gga Gly		_	_	1299
_						-	_	-	_		_	tgg Trp	-			1347
												aat Asn				1395
att Ile	tat Tyr	caa Gln 465	aca Thr	cac His	aag Lys	agg Arg	gtg Val 470	tgg Trp	cct Pro	gct Ala	tct Ser	cag Gln 475	cga Arg	gac Asp	gta Val	1443
												act Thr				1491
											Asp	cat His				1539
					Cys					Ile		gtt Val				1587
				Val					Gly					Ser	agg	1635
			Leu					Tyr					Asn		gga Gly	1683
gga Gly	tgg Trp 560	Ala	cca Pro	geo Ala	tca Ser	gtg Val	. Leu	agg Arg	gca Ala	gtg Val	gca Ala 570	r ras	cga Arg	gag Glu	tat Tyr	1731

cct aaa ttt cta aaa cgt ttt act tct tac gtc caa gaa aaa act gca 1779
Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln Glu Lys Thr Ala
575 580 585 590

gga aag oot att ttg tto tagtattaac aggtactaga agatatgttt 1827 Gly Lys Pro Ile Leu Phe 595

tatctttttt taactttatt tgactaatat gactgtcaat actaaaattt agttgttgaa 1887

agtatttact atgttttttc cggaattc 1915

<210> 22

<211> 596

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: FLAG-GPBPDSXY/NLS

<400> 22

Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met Ser Asp 10 15

Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu Thr Glu
20 25 30

Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn 35 40 45

Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala 50 55 60

Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly 65 70 75 80

Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu 85 90 95

Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu Arg Ala 100 105 110

Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu Gln His 115 120 125

Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg His Gly
130 135 140

Lys Gly His Ser Leu Arg Glu Lys Leu Ala Glu Met Glu Thr Phe Arg 145 150 155 160

Asp Ile Leu Cys Arg Gln Val Asp Thr Leu Gln Lys Tyr Phe Asp Ala 165 170 175

Cys Ala Asp Ala Val Ser Lys Asp Glu Leu Gln Arg Asp Lys Val Val
180 185 190

Glu Asp Asp Glu Asp Asp Phe Pro Thr Thr Arg Ser Asp Gly Asp Phe 195 200 205

48

Leu His Ser Thr Asn Gly Asn Lys Glu Lys Leu Phe Pro His Val Thr 210 215 220

- Pro Lys Gly Ile Asn Gly Ile Asp Phe Lys Gly Glu Ala Ile Thr Phe 225 235 240
- Lys Ala Thr Thr Ala Gly Ile Leu Ala Thr Leu Ser His Cys Ile Glu 245 250 255
- Leu Met Val Lys Arg Glu Asp Ser Trp Gln Lys Arg Leu Asp Lys Glu 260 265 270
- Thr Glu His Phe Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu 275 280 285
- Ile Asn Glu Glu Glu Phe Phe Asp Ala Val Glu Ala Ala Leu Asp Arg 290 295 300
- Gln Asp Lys Ile Glu Glu Gln Ser Gln Ser Glu Lys Val Arg Leu His 305 310 315 320
- Trp Pro Thr Ser Leu Pro Ser Gly Asp Ala Phe Ser Ser Val Gly Thr 325 330 335
- His Arg Phe Val Gln Lys Pro Tyr Ser Arg Ser Ser Ser Met Ser Ser 340 345 350
- Ile Asp Leu Val Ser Ala Ser Asp Asp Val His Arg Phe Ser Ser Gln 355 360 365
- Val Glu Glu Met Val Gln Asn His Met Thr Tyr Ser Leu Gln Asp Val 370 375 380
- Gly Gly Asp Ala Asn Trp Gln Leu Val Val Glu Glu Gly Glu Met Lys 385 390 395 395
- Val Tyr Arg Arg Glu Val Glu Glu Asn Gly Ile Val Leu Asp Pro Leu 405 410 415
- Lys Ala Thr His Ala Val Lys Gly Val Thr Gly His Glu Val Cys Asn 420 425 430
- Tyr Phe Trp Asn Val Asp Val Arg Asn Asp Trp Glu Thr Thr Ile Glu 435 440 445
- Asn Phe His Val Val Glu Thr Leu Ala Asp Asn Ala Ile Ile Ile Tyr 450 455 460
- Gln Thr His Lys Arg Val Trp Pro Ala Ser Gln Arg Asp Val Leu Tyr 465 470 475 480
- Leu Ser Val Ile Arg Lys Ile Pro Ala Leu Thr Glu Asn Asp Pro Glu 485 490 495
- Thr Trp Ile Val Cys Asn Phe Ser Val Asp His Asp Ser Ala Pro Leu 500 505 510
- Asn Asn Arg Cys Val Arg Ala Lys Ile Asn Val Ala Met Ile Cys Gln 515 520 525

Thr Leu Val Ser Pro Pro Glu Gly Asn Gln Glu Ile Ser Arg Asp Asn 535 Ile Leu Cys Lys Ile Thr Tyr Val Ala Asn Val Asn Pro Gly Gly Trp 550 555 Ala Pro Ala Ser Val Leu Arg Ala Val Ala Lys Arg Glu Tyr Pro Lys Phe Leu Lys Arg Phe Thr Ser Tyr Val Gln Glu Lys Thr Ala Gly Lys 585 Pro Ile Leu Phe 595 <210> 23 <211> 2038 <212> DNA <213> Artificial Sequence <223> Description of Artificial Sequence: GPBP-D169A <220> <221> CDS <222> (10)..(1920) <400> 23 gaattcacc atg gcc cca cta gcc gac tac aag gac gac gat gac aag atg 51 Met Ala Pro Leu Ala Asp Tyr Lys Asp Asp Asp Lys Met 1 10 tog gat aat cag ago tgg aac tog tog ggc tog gag gag gat coa gag Ser Asp Asn Gln Ser Trp Asn Ser Ser Gly Ser Glu Glu Asp Pro Glu 20 acg gag tot ggg ccg cct gtg gag cgc tgc ggg gtc ctc agt aag tgg Thr Glu Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp aca aac tac att cat ggg tgg cag gat cgt tgg gta gtt ttg aaa aat 195 Thr Asn Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn 55 aat gct ctg agt tac tac aaa tct gaa gat gaa aca gag tat ggc tgc 243 Asn Ala Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys aga gga tcc atc tgt ctt agc aag gct gtc atc aca cct cac gat ttt Arg Gly Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe gat gaa tgt cga ttt gat att agt gta aat gat agt gtt tgg tat ctt 339 Asp Glu Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu 95 100 cgt gct cag gat cca gat cat aga cag caa tgg ata gat gcc att gaa 387 Arg Ala Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu 115 120

cag Gln	cac His	aag Lys	act Thr 130	g aa Glu	tct Ser	gga Gly	tat Tyr	gga Gly 135	tct Ser	gaa Glu	tcc Ser	agc Ser	ttg Leu 140	cgt Arg	cga Arg	435
cat His	ggc Gly	tca Ser 145	atg Met	gtg Val	tcc Ser	ctg Leu	gtg Val 150	tct Ser	gga Gly	gca Ala	agt Ser	ggc Gly 155	tac Tyr	tct Ser	gca Ala	483
aca Thr	tcc Ser 160	acc Thr	tct Ser	tca Ser	ttc Phe	aag Lys 165	aaa Lys	ggc Gly	cac His	agt Ser	tta Leu 170	cgt Arg	gag Glu	aag Lys	ttg Leu	531
gct Ala 175	gaa Glu	atg Met	g aa Glu	aca Thr	ttt Phe 180	aga Arg	gcc Ala	atc Ile	tta Leu	tgt Cys 185	aga Arg	caa Gln	gtt Val	gac Asp	acg Thr 190	579
cta Leu	cag Gln	aag Lys	tac Tyr	ttt Phe 195	gat Asp	gcc Ala	tgt Cys	gct Ala	gat Asp 200	gct Ala	gtc Val	tct Ser	aag Lys	gat Asp 205	gaa Glu	627
ctt Leu	caa Gln	agg Arg	gat Asp 210	aaa Lys	gtg Val	gta Val	gaa Glu	gat Asp 215	gat Asp	gaa Glu	gat Asp	gac Asp	ttt Phe 220	cct Pro	aca Thr	675
acg Thr	cgt Arg	tct Ser 225	gat Asp	ggt Gly	gac Asp	ttc Phe	ttg Leu 230	cat His	agt Ser	acc Thr	aac Asn	ggc Gly 235	aat Asn	aaa Lys	gaa Glu	723
aag Lys	tta Leu 240	Phe	cca Pro	cat His	gtg Val	aca Thr 245	cca Pro	aaa Lys	gga Gly	att Ile	aat Asn 250	ggt	ata Ile	gac Asp	ttt Phe	771
aaa Lys 255	Gly	gaa Glu	gcg Ala	ata Ile	act Thr 260	Phe	aaa Lys	gca Ala	act Thr	act Thr 265	Ala	gga Gly	atc Ile	ctt Leu	gca Ala 270	819
aca Thr	ctt Leu	tct Ser	cat His	tgt Cys 275	Ile	gaa Glu	cta Leu	atg Met	gtt Val 280	Lys	cgt Arg	gag Glu	gac Asp	e ago Ser 285	tgg Trp	867
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gca Ala	a tat a Tyr	aaa Ly: 30!	a Ası	t gca n Ala	atg Met	aca Thr	gaa Glu 310	ı Lev	: aaçı Lys	g aaa Lys	a aaa s Lys	s Ser	r His	ttt Phe	gga Gly	963
gg:	a cca y Pro 32	As	t ta p Ty:	t gaa r Gli	a gaa 1 Glu	a ggo 1 Gly 325	Pro	t aad o Asi	agt Sei	c Ctq	g att u Ile 330	e Ası	t gaa n Gli	a gaa u Gli	a gag u Glu	1011
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aca Thr	tat Tyr	gta Val	gct Ala	aat Asn 595	gtg Val	aac Asn	cct Pro	gga Gly	gga Gly 600	tgg Trp	gca Ala	cca Pro	gcc Ala	tca Ser 605	gtg Val	1827
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1920

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625 630 635

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Ser Gly Pro Pro Val Glu Arg Cys Gly Val Leu Ser Lys Trp Thr Asn 35 40 45

Tyr Ile His Gly Trp Gln Asp Arg Trp Val Val Leu Lys Asn Asn Ala 50 55 60

Leu Ser Tyr Tyr Lys Ser Glu Asp Glu Thr Glu Tyr Gly Cys Arg Gly 65 70 75 80

Ser Ile Cys Leu Ser Lys Ala Val Ile Thr Pro His Asp Phe Asp Glu 85 90 95

Cys Arg Phe Asp Ile Ser Val Asn Asp Ser Val Trp Tyr Leu Arg Ala

Gln Asp Pro Asp His Arg Gln Gln Trp Ile Asp Ala Ile Glu Gln His
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Lys Thr Glu Ser Gly Tyr Gly Ser Glu Ser Ser Leu Arg Arg His Gly 130 135 140

Ser Met Val Ser Leu Val Ser Gly Ala Ser Gly Tyr Ser Ala Thr Ser 145 150 155 160

Thr Ser Ser Phe Lys Lys Gly His Ser Leu Arg Glu Lys Leu Ala Glu 165 170 175

Met Glu Thr Phe Arg Ala Ile Leu Cys Arg Gln Val Asp Thr Leu Gln
180 185 190

Lys Tyr Phe Asp Ala Cys Ala Asp Ala Val Ser Lys Asp Glu Leu Gln
195 200 205

Arg Asp Lys Val Val Glu Asp Asp Glu Asp Asp Phe Pro Thr Thr Arg

PCT/IB00/00324 WO 00/50607

Ser Asp Gly Asp Phe Leu His Ser Thr Asn Gly Asn Lys Glu Lys Leu 235 230 Phe Pro His Val Thr Pro Lys Gly Ile Asn Gly Ile Asp Phe Lys Gly Glu Ala Ile Thr Phe Lys Ala Thr Thr Ala Gly Ile Leu Ala Thr Leu 265 Ser His Cys Ile Glu Leu Met Val Lys Arg Glu Asp Ser Trp Gln Lys Arg Leu Asp Lys Glu Thr Glu Lys Lys Arg Arg Thr Glu Glu Ala Tyr 295 Lys Asn Ala Met Thr Glu Leu Lys Lys Ser His Phe Gly Gly Pro Asp Tyr Glu Glu Gly Pro Asn Ser Leu Ile Asn Glu Glu Glu Phe Phe 330 Asp Ala Val Glu Ala Ala Leu Asp Arg Gln Asp Lys Ile Glu Glu Gln

- 345
- Ser Gln Ser Glu Lys Val Arg Leu His Trp Pro Thr Ser Leu Pro Ser 360
- Gly Asp Ala Phe Ser Ser Val Gly Thr His Arg Phe Val Gln Lys Pro
- Tyr Ser Arg Ser Ser Ser Met Ser Ser Ile Asp Leu Val Ser Ala Ser 390
- Asp Asp Val His Arg Phe Ser Ser Gln Val Glu Met Val Gln Asn
- His Met Thr Tyr Ser Leu Gln Asp Val Gly Gly Asp Ala Asn Trp Gln 425
- Leu Val Val Glu Glu Gly Glu Met Lys Val Tyr Arg Arg Glu Val Glu
- Glu Asn Gly Ile Val Leu Asp Pro Leu Lys Ala Thr His Ala Val Lys
- Gly Val Thr Gly His Glu Val Cys Asn Tyr Phe Trp Asn Val Asp Val 475 465
- Arg Asn Asp Trp Glu Thr Thr Ile Glu Asn Phe His Val Val Glu Thr 485
- Leu Ala Asp Asn Ala Ile Ile Ile Tyr Gln Thr His Lys Arg Val Trp 505
- Pro Ala Ser Gln Arg Asp Val Leu Tyr Leu Ser Val Ile Arg Lys Ile 520
- Pro Ala Leu Thr Glu Asn Asp Pro Glu Thr Trp Ile Val Cys Asn Phe 535 530

Ser Val Asp His Asp Ser Ala Pro Leu Asn Asn Arg Cys Val Arg Ala 545 550 560

Lys Ile Asn Val Ala Met Ile Cys Gln Thr Leu Val Ser Pro Pro Glu 565 570 575

Gly Asn Gln Glu Ile Ser Arg Asp Asn Ile Leu Cys Lys Ile Thr Tyr 580 585 590

Val Ala Asn Val Asn Pro Gly Gly Trp Ala Pro Ala Ser Val Leu Arg 595 600 605

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Thr Arg Gly Phe Val Phe Thr Arg His Ser Gln Thr Thr Ala Ile Pro
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tca tgt cca gag ggg aca gtg cca ctc tac agt ggg ttt tct ttt ctt
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Ser Cys Pro Glu Gly Thr Val Pro Leu Tyr Ser Gly Phe Ser Phe Leu
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Phe Val Gln Gly Asn Gln Arg Ala His Gly Gln Asp Leu Gly Thr Leu
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Gly Ser Cys Leu Gln Arg Phe Thr Thr Met Pro Phe Leu Phe Cys Asn
gtc aat gat gta tgt aat ttt gca tct cga aat gat tat tca tac tgg
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Val Asn Asp Val Cys Asn Phe Ala Ser Arg Asn Asp Tyr Ser Tyr Trp
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Leu Ser Thr Pro Ala Leu Met Pro Met Asn Met Ala Pro Ile Thr Gly
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Val	Asn	Asp	Val	Суs 85	Asn	Phe	Ala	Ser	Arg 90	Asn	Asp	Tyr	Ser	Tyr 95	Trp	
Leu	Ser	Thr	Pro	Ala	Leu	Met	Pro	Met	Asn	Met	Ala	Pro	Ile	Thr	Gly	

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Ser Cys Pro Glu Gly Thr Val Pro Leu Tyr Ser Gly Phe Ser Phe Leu

PCT/IB00/00324

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acg aga ggc ttt gtc ttc acc cga cac agt caa acc aca gca att cct Thr Arg Gly Phe Val Phe Thr Arg His Ser Gln Thr Thr Ala Ile Pro

20 25 30 tca tgt cca gag ggg aca gtg cca ctc tac agt ggg ttt tct ttt ctt 144 Ser Cys Pro Glu Gly Thr Val Pro Leu Tyr Ser Gly Phe Ser Phe Leu 35 40 ttt gta caa gga aat caa cga gcc cac gga caa gac ctt gat gca ctg 192 Phe Val Gln Gly Asn Gln Arg Ala His Gly Gln Asp Leu Asp Ala Leu 55 ttt gtg aag gto ctg cga tcg cca tagccgttca cagccaaacc actgacattc 246 Phe Val Lys Val Leu Arg Ser Pro ctccatgtcc tcacggctgg atttctctct ggaaaggatt ttcattcatc atgaaagcct 306 attocatoaa otgtgaaago tggggaatta gaaaaaataa taagtogotg toaggtgtgo 366 atgaagaaaa gacactgaag ctaaaaaaaga cagcagaact gctatttttc atcctaaaga 426 acaaaqtaat gacagaacat gctgttattt aggtattttt ctttaaccaa acaatattgc 486 507 tccatgatga cttagtacaa a <210> 52 <211> 72 <212> PRT <213> Artificial Sequence <220> <223> Description of Artificial Sequence: GPDIII-V <400> 52 Gly Leu Lys Gly Lys Arg Gly Asp Ser Gly Ser Pro Ala Thr Trp Thr Thr Arg Gly Phe Val Phe Thr Arg His Ser Gln Thr Thr Ala Ile Pro Ser Cys Pro Glu Gly Thr Val Pro Leu Tyr Ser Gly Phe Ser Phe Leu Phe Val Gln Gly Asn Gln Arg Ala His Gly Gln Asp Leu Asp Ala Leu 55 Phe Val Lys Val Leu Arg Ser Pro <210> 53 <211> 659 <212> DNA <213> Artificial Sequence <220> <223> Description of Artificial Sequence: HMBP-21 <220>

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ccc Pro	tcc Ser	cag Gln	agg Arg 10	cac His	gga Gly	tcc Ser	aag Lys	tac Tyr 15	ctg Leu	gcc Ala	aca Thr	gca Ala	agt Ser 20	acc Thr	atg Met	102
gac Asp	cat His	gcc Ala 25	agg Arg	cat His	ggc	ttc Phe	ctc Leu 30	cca Pro	agg Arg	cac His	aga Arg	gac Asp 35	acg Thr	ggc Gly	atc Ile	150
ctt Leu	gac Asp 40	tcc Ser	atc Ile	999 Gly	egc Arg	ttc Phe 45	ttt Phe	ggc Gly	ggt Gly	gac Asp	agg Arg 50	ggt Gly	gcg Ala	cca Pro	aag Lys	198
cgg Arg 55	ggc Gly	tct Ser	ggc Gly	aag Lys	gta Val 60	ccc Pro	tgg Trp	cta Leu	aag Lys	ccg Pro 65	ggc Gly	cgg Arg	agc Ser	cct Pro	ctg Leu 70	246
ccc Pro	tct Ser	cat His	gcc Ala	cgc Arg 75	agc Ser	cag Gln	cct Pro	ggg gly	ctg Leu 80	tgc Cys	aac Asn	atg Met	tac Tyr	aag Lys 85	gac Asp	294
tca Ser	cac His	cac His	ccg Pro 90	gca Ala	aga Arg	act Thr	gct Ala	cac His 95	tat Tyr	ggc Gly	tcc Ser	ctg Leu	ccc Pro 100	cag Gln	aag Lys	342
tca Ser	cac His	ggc Gly 105	cgg Arg	acc Thr	caa Gln	gat Asp	gaa Glu 110	aac Asn	ccc Pro	gta Val	gtc Val	cac His 115	ttc Phe	ttc Phe	aag Lys	390
aac Asn	att Ile 120	gtg Val	acg Thr	cct Pro	cgc Arg	aca Thr 125	cca Pro	ccc Pro	ccg Pro	tcg Ser	cag Gln 130	gga Gly	aag Lys	ggg ggg	aga Arg	438
gga Gly 135	ctg Leu	tcc Ser	ctg Leu	agc Ser	aga Arg 140	ttt Phe	agc Ser	tgg Trp	Gly aaa	gcc Ala 145	gaa Glu	ggc	cag Gln	aga Arg	cca Pro 150	486
gga Gly	ttt Phe	ggc Gly	tac Tyr	gga Gly 155	ggc Gly	aga Arg	gcg Ala	tcc Ser	gac Asp 160	tat Tyr	aaa Lys	tcg Ser	gct Ala	cac His 165	aag Lys	534
gga Gly	ttc Phe	aag Lys	gga Gly 170	gtc Val	gat Asp	gcc Ala	cag Gln	ggc Gly 175	acg Thr	ctt Leu	tcc Ser	aaa Lys	att Ile 180	ttt Phe	aag Lys	582
ctg Leu	gga Gly	gga Gly 185	aga Arg	gat Asp	agt Ser	cgc Arg	tct Ser 190	gga Gly	tca Ser	ccc Pro	atg Met	gct Ala 195	aga Arg	cgc Arg		627
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His Arg Asp Thr Gly Ile Leu Asp Ser Ile Gly Arg Phe Phe Gly Gly
35 40 45

Asp Arg Gly Ala Pro Lys Arg Gly Ser Gly Lys Val Pro Trp Leu Lys
50 55 60

Pro Gly Arg Ser Pro Leu Pro Ser His Ala Arg Ser Gln Pro Gly Leu
65 70 75 80

Cys Asn Met Tyr Lys Asp Ser His His Pro Ala Arg Thr Ala His Tyr 85 90 95

Gly Ser Leu Pro Gln Lys Ser His Gly Arg Thr Gln Asp Glu Asn Pro 100 105 110

Val Val His Phe Phe Lys Asn Ile Val Thr Pro Arg Thr Pro Pro Pro 115 120 125

Ser Gln Gly Lys Gly Arg Gly Leu Ser Leu Ser Arg Phe Ser Trp Gly
130 135 140

Ala Glu Gly Gln Arg Pro Gly Phe Gly Tyr Gly Gly Arg Ala Ser Asp 145 150 155 160

Tyr Lys Ser Ala His Lys Gly Phe Lys Gly Val Asp Ala Gln Gly Thr 165 170 175

Leu Ser Lys Ile Phe Lys Leu Gly Gly Arg Asp Ser Arg Ser Gly Ser 180 185 190

Pro Met Ala Arg Arg 195